TITLE OF THE INVENTION

HUMAN SEMAPHORIN L (H-SEMAL) AND CORRESPONDING SEMAPHORINS IN OTHER SPECIES

RELATED APPLICATIONS

This application claims priority to German Application Nos. 19729211.9 and 19805371.1, filed July 9, 1997 and February 11, 1998 respectively, each incorporated herein by reference.

BACKGROUND OF THE INVENTION

Field of the Invention

The invention relates to novel semaphorins which are distinguished by a particular domain structure and derivatives thereof, nucleic acids (DNA, RNA, cDNA) which code for these semaphorins, and derivatives thereof, and the preparation and use thereof.

Description of the Related Art

The publications which are referenced in this application describe the state of the art to which this invention pertains. These references are incorporated herein by references.

Semaphorins were described for the first time by Kolodkin {Kolodkin et al. (1993) Cell 75:1389-1399} as members of a conserved gene family.

The genes or parts of the genes of other semaphorins have now been cloned and, in some cases, characterized. To date, a total of 5 human (H-Sema III, H-Sema V, H-Sema IV, H-SemaB and H-SemaE) {Kolodkin et al. (1993); Roche et al. (1996) Onkogene 12:1289-1297; Sekido et al. (1996) Proc. Natl.

Acad. Sci. USA 93:4120-4125; Xiang et al. (1996) Genomics 32:39-48; Hall et al. (1996) Proc. Natl. Acad. Sci. USA 39:11780-11785; Yamada et al. (1997) (GenBank Accession No. AB000220)}, 8 murine (mouse genes; M-Sema A to M-Sema-H) {Püschel et al. (1995) Neuron 14:941-948; Messerschmidt et al. (1995) Neuron 14:949-959; Inigaki et al. (1995) FEBS Letters 370:269-272; Adams et al. (1996) Mech. Dev. 57:33-45; Christensen et al. (1996)(GenBank Accession No. Z80941, Z93948)}, 5 galline (chicken) (collapsin-1 to -5) {Luo et al. (1993); Luo et al. (1995) Neuron 14:1131-1140}, and genes from rats (R-Sema-III) {Giger et al. (1996) J. Comp. Neurol. 375:378-392}, zebra fish, insects (fruit fly (Drosophila melanogaster: D-Sema I and D-Sema II), beetles (Tribolium confusum: T-Sema-I), grasshoppers (Schistocerca americana: G-Sema-I)) {Kolodkin et al. (1993)}, and nematodes (C.elegans: Ce-Sema) {Roy et al. (1994) (GenBank Accession No. U15667)} have been disclosed. In addition, two poxviruses (vaccinia (ORF-A39) and variola (ORFA39-homologous)) {Kolodkin et al. (1993)} and alcelaphine herpesvirus Type 1 (AHV-1) (AHV-Sema) {Ensser and Fleckenstein (1995) Gen. Virol. 76:1063-1067} have genes homologous to semaphorins.

Table 1 summarizes the semaphorins identified to date in various species. Table 1 indicates the names of the semaphorins (column 1), the synonyms used (column 2), the species from which the particular semaphorin has been isolated (column 3) and, where known, data on the domain structure of the encoded protein and on the chromosomal location (column 4 in Table 1), the accession number under which the sequence of the gene is stored in gene databanks (for example in an EST (expressed sequence tags) databank, EMBL (European Molecular Biology Laboratory, Heidelberg) or NCBI (National Center for Biotechnology Information, Maryland, USA), and the corresponding reference under which these data have been published (column 5 in Table 1).

All the gene products (encoded semaphorins) of the semaphorin genes disclosed to date have an N-terminal signal peptide which has at its C-terminal end a characteristic Sema domain with a length of about 450 to 500 amino acids. Highly conserved amino acid motifs and a number of highly

conserved cysteine residues are located within the Sema domains. The gene products (semaphorins) differ in the C-terminal sequences which follow the Sema domains and are composed of one or more domains. They have, for example, in these C-terminal amino acid sequences transmembrane domains (TM), immunoglobulin-like domains (Ig) (constant part of the immunoglobulin), cytoplasmic sequences (CP), processing signals (P) (for example having the consensus sequence (RXR) where R is the amino acid arginine and X is any amino acid) and/or hydrophilic C termini (HPC). The semaphorins disclosed to date can be divided on the basis of the differences in the domain structure in the C terminus into 5 different subgroups (I to V):

I		Secreted, without other domains (for example ORF-A49)
II	Ig	Secreted (without transmembrane domain) for example AHV-Sema)
III	Ig, TM, CP	Membrane-anchored with cytoplasmic sequence (for example CD100)
IV	Ig, (P), HPC	Secreted with hydrophilic C terminus (for example H-Sema III, M-SemaD, collapsin-1)
V	Ig, TM, CP	Membrane-anchored with C-terminal 7 thrombospondin motif (for example M-SemaF and G)

A receptor or extracellular ligand for semaphorins has not been described to date. Intracellular, heterotrimeric GTP-binding protein complexes have been described in connection with semaphorin-mediated effects. One component of these protein complexes which has been identified in chickens is called CRMP (collapsin response mediator protein) and is presumed to be a component of the semaphorin-induced intracellular signal cascade (Goshima et al. (1995) Nature 376: 509-514). CRMP62, for example, has homology with unc-33, a nematode protein which is essential for directed growth of axons. A human protein with 98% amino acid identity with CRMP62 is likewise known (Hamajima et al. (1996) Gene 180: 157-163). Several CRMP-related genes have likewise been described in rats (Wang et al. (1996) Neurosci. 16: 6197-6207).

The secreted or transmembrane semaphorins convey repulsive signals for growing nerve buds. They play a part in the development of the central nervous system (CNS) and are expressed in particular in muscle and nerve tissues (Kolodkin et al. (1993); Luo et al. (1993) Cell 75:217-227).

Pronounced expression of M-SemaG has been observed not only in the CNS but also in cells of the lymphatic and hematopoietic systems, in contrast to the closely related M-SemaF {Furuyima et al. (1996) J. Biol. Chem. 271: 33376-33381}.

Recently, two other human semaphorins have been identified, H-Sema IV and H-Sema V, specifically in a region on chromosome 3p21.3, whose deletion is associated with various types of bronchial carcinomas. H-Sema IV {Roche et al. (1996), Xiang et al. (1996), Sekido et al. (1996)} is about 50% identical at the amino acid level with M-SemaE, whereas H-Sema V {Sekido et al. (1996)} is the direct homolog of M-SemaA (86% amino acid identity). Since these genes (H-Sema IV and V) were found during DNA sequencing projects on the deleted 3p21.3 loci, the complex intron-exon structure of these two genes is known. Both genes are expressed in various neuronal and non-neuronal tissues.

Likewise only recently, the cellular surface molecule CD100 (human), expressed and induced on activated T cells, has been identified as a semaphorin (likewise listed in Table 1). It assists interaction with B cells via the CD40 receptor and the corresponding ligand CD40L. CD100 is a membrane-anchored glycoprotein dimer of 150 kd (kilodaltons). An association of the intracytoplasmic C-terminus of CD100 with an as yet unknown kinase has been described {Hall et al. (1996)}. This means that CD100 is the first and to date only semaphorin whose expression in cells of the immune system has been demonstrated.

In the "transforming genes of rhadinoviruses" project, the complete genome of alcelaphine herpesvirus Type 1 (AHV-1) has been cloned and sequenced {Ensser et al. (1995)}. AHV-1 is the causative agent of malignant catarrhal fever, a disease of various ruminants which is associated with a lymphoproliferative syndrome and is usually fatal. On analysis, an open reading frame was found, at one end of the viral genome, having remote but significant homology with a gene of vaccinia- virus (ORF-A39 corresponds to VAC-A39 in Ensser et al. (1995) J. Gen. Virol. 76:1063-1067) which has been assigned to the semaphorin gene family. Whereas the AHV-1 semaphorin (AHV-Sema) has a well-conserved semaphorin structure, the poxvirus genes (ORF-A39 and ORF-A39-homologous, see Table 1) have C-terminal truncations, i.e. the conserved Sema domain is present in them only incompletely.

Databank comparison of the found AHV-Sema with dbEST (EST (expressed sequence tags) databank (db)) provided in each case 2 EST sequences from 2 independent cDNA clones from human placenta (accession numbers H02902, H03806 (clone 151129), accession numbers R33439 and R33537 (clone 135941)). These display distinctly greater homology with AHV-1 semaphorin than with the neuronal semaphorins hitherto described.

SUMMARY OF THE INVENTION

The present invention relates to semaphorins which have a novel, as yet undisclosed and unexpected domain structure and which possess a biochemical function in the immune system (immunomodulating semaphorins). The novel semaphorins are referred to as type L semaphorins (SemaL). They comprise an N-terminal signal peptide, a characteristic Sema domain and, in the C-terminal region of the protein, an immunoglobulin-like domain and a hydrophobic domain which represents a potential transmembrane domain.

The amino acid sequence of the signal peptide may have fewer than 70, preferably fewer than 60 amino acids and more than 20, preferably more than 30 amino acids, and a particularly preferred length is of about 40 to 50 amino acids. In a specific embodiment of the invention, the signal peptide has a length of 44 amino acids, i.e. a cleavage site for a signal peptidase is located between amino acids 44 and 45.

The Sema domain may have a length of from 300 to 700 or more, preferably of about 400 to 600, amino acids. Preferred Sema domains have a length of 450 to 550 amino acids, preferably of about 500 amino acids. In a preferred embodiment of the invention, the Sema domain is joined to the signal peptide, in which case the Sema domain preferably extends up to amino acid 545.

The immunoglobulin-like domain may have a length of about 30 to 110 or more amino acids, and preferred lengths are between 50 and 90, particularly preferably about 70, amino acids.

The transmembrane domain may have a length of about 10 to 35, preferably of about 15 to 30, particularly preferably of about 20 to 25, amino acids.

The invention relates to type L semaphorins from various species, in particular from vertebrates, for example from birds and/or fishes, preferably from mammals, for example from primates, rat, rabbit, dog, cat, sheep, goat, cow, horse, pig, particularly preferably from human and mouse. The invention also relates to corresponding semaphorins from microorganisms, especially from pathogenic microorganisms, for example from bacteria, yeasts and/or viruses, for example from retroviruses, especially from human-pathogenic microorganisms.

BRIEF DECEPTION OF THE DRAWING

The invention will be described in greater detail with the aid of the following figures:

Fig. 1 is a Multiple tissue Northern blot for the tissue-specific expression of H-SemaL.

Fig. 2 is a diagrammic representation of the cloning of the H-SemaL cDNA and of the genomic organization of the H-SemaL encoding sequence.

Fig. 3 is a phylogenetic tree.

Fig. 4 is a FACS analysis of H-SEMAL expression in various cell lines.

Fig. 5 is a comparative analysis of CD 100 and H-SemaL expression.

Fig. 6 is the expression of secretable human SEMA-L (H-SemaL) in HiFive and SC3 cells.

Fig. 7 depicts the specificity of the antiserum.

Fig. 8 is a plasmid map of pMelBacA-H-SEMAL.

DETAILED DESCRIPTION OF THE INVENTION

One embodiment of the invention is a corresponding human semaphorin (H-SemaL) which has a signal peptide, a Sema domain, an immunoglobulin-like domain and a transmembrane domain. A specific embodiment is the semaphorin which is given by the amino acid sequence shown in Table 4.

Another embodiment of the invention comprises corresponding semaphorins in other species which have, in the region of the Sema domain, an amino acid identity greater than 40%, preferably greater than 50%, particularly preferably greater than 60%, in relation to the Sema domain of H-SemaL (amino acids 45 to 545 of the sequence in Table 4). The corresponding semaphorins from closely related species (for example primates, mouse) may perfectly well have

amino acid identities of greater than 70%, preferably greater than 80%, particularly preferably greater than 90%. Percentage homologies can be determined or calculated for example using the GAP program (GCG program package, Genetic Computer Group (1991)).

Such an embodiment of the invention is a corresponding mouse semaphorin (murine semaphorin (M-SemaL)). This contains, for example, the partial amino acid sequence shown in Table 5 (murine semaphorin (M-SemaL)).

The invention also relates to corresponding semaphorins which have an amino acid identity (considered over the entire length of the amino acid sequence of the protein) of only about 15 to 20% in the case of less related species (very remote from one another phylogenetically), preferably 25 to 30%, particularly preferably 35 to 40%, or a higher identity in relation to the complete amino acid sequence of H-SemaL shown in Table 4.

The genes which code for type L semaphorins have a complex exon-intron structure. These genes may have, for example, between 10 and 20 exons, preferably about 11 to 18, particularly preferably 12 to 16, exons and a corresponding number of introns. However, they may also have the same number of exons and introns as does the gene of H-SemaL (13 or 15 exons, preferably 14 exons). A particular embodiment of the invention relates to the gene of H-SemaL. This gene preferably has a length of 8888 to 10,000 or more nucleotides. The human semaphorin gene preferably contains the nucleotide sequence given in Table 14 or the nucleotide sequence which has been deposited at the GenBank® databank under accession number AF030697. These nucleotide sequences contain at least 13 introns. In addition, the human semaphorin gene has at the 5' end an additional sequence region. This region contains, where appropriate, further coding and uncoding sequences, for example one or two further introns or exons.

Attempts to locate the human type L semaphorin on the chromosome revealed that the corresponding gene is located at position 15q22.3-23. The gene for M-SemaL has correspondingly been located at position 9A3.3-B.

As a consequence of the complex intron-exon structure, the splicing of the primary transcript of the semaphorin mRNA may vary, resulting in different splicing variants of the semaphorins. The proteins translated from these splicing variants are derivatives of the semaphorins according to the invention. They correspond in their amino acid sequence and also substantially in their domain structure to the described type L semaphorins according to the invention, but are truncated by comparison with the latter where appropriate. For example, splicing variants wholly or partly lacking the transmembrane domain may be formed. A semaphorin derivative which contains an incomplete, or no, transmembrane domain, but contains a signal peptide, may be secreted and in this way have effects outside the cell, locally or else over relatively large distances, for example on other cells. Another splicing variant may, for example, no longer contain a sequence which codes for a signal peptide and, where appropriate, also no sequence which codes for a hydrophobic amino acid sequence representing a potential transmembrane domain. One consequence would be that this semaphorin derivative is neither incorporated into the membrane nor secreted (unless through secretory vesicles). Such a semaphorin derivative may be involved in intracellular processes, for example in signal transduction processes. It is possible in this way for a wide variety of intra- and extracellular processes to be controlled and/or harmonized with the same basic molecule (type L semaphorins) and the derivatives derived therefrom (for example splicing variants).

A particular embodiment of the invention relates to semaphorin derivatives which are derived from the type L semaphorins according to the invention but which contain an incomplete, or no, transmembrane domain.

Another embodiment of the invention relates to semaphorin derivatives which are derived from the type L semaphorins according to the invention but which contain no signal peptide.

The signal peptide may also undergo post-translational elimination. This forms a membrane-bound (with TM domain) or a secreted (splicing variant without TM domain) semaphorin derivative with truncated domain structure. A semaphorin derivative which has undergone post-translational processing in this way now contains only Sema domain, Ig domain and, where appropriate, transmembrane domain. A signal peptide cleavage site can be located, for example, right at the end of the signal peptide, but it may, for example, be located 40 to 50 amino acids or more away from the amino terminus.

A "truncated" (i.e. containing fewer domains) semaphorin L derivative can be distinguished from other semaphorins which are not derived from type L semaphorins in that there is a very great (> 90%) amino acid identity or an identical amino acid sequence with the type L semaphorins in the domains which are present.

The semaphorins according to the invention may also have undergone post-translational modification in other ways. For example, they may be glycosylated (N- and/or O-glycosylated) once, twice, three, four, five, six, seven, eight, nine, ten or more times. The amino acid sequences of the semaphorins may then have an equal number of or more consensus sequences for potential glycosylation sites, preferably five such sites. One embodiment of the invention relates to semaphorins in which the glycosylation sites are located at positions which correspond to positions 105, 157, 258, 330 and 602 of the H-SemaL amino acid sequence (Table 4).

In addition, the semaphorins may be in the form of their phosphorylated derivatives. Semaphorins may be the substrates of various kinases, for example the amino acid sequences may have consensus sequences for protein kinase C, tyrosine kinase and/or creatine kinases. In addition, the

amino acid sequences of the semaphorins may have consensus sequences for potential myristylation sites. Corresponding semaphorin derivatives may be esterified with myristic acid at these sites.

The type L semaphorins according to the invention and their derivatives may be in the form of monomers, dimers and/or multimers, for example two or more semaphorins or their derivatives can be linked together by intermolecular disulfide bridges. It is also possible for intramolecular disulfide bridges to be formed.

Further derivatives of the semaphorins according to the invention are fusion proteins. A fusion protein of this type contains, on the one hand, a type L semaphorin or parts thereof and, in addition, another peptide or protein or a part thereof. Peptides or proteins or parts thereof may be, for example, epitope tags (for example His tag (6xhistidine), Myc tag, flu tag) which can be used, for example, for purifying the fusion proteins, or those which can be used for labeling the fusion proteins, for example GFP (green fluorescent protein). Examples of derivatives of the type L semaphorins are given for example by the constructs described in the examples. The sequences of these constructs can be found in Tables 7 to 15, where appropriate taking account of the annotations relating to the plasmids.

The invention further relates to nucleic acid sequences, preferably DNA and RNA sequences, which code for the type L semaphorins according to the invention and/or their derivatives, for example the corresponding genes, the various splicing variants of the mRNA, the cDNAs corresponding thereto, and derivatives thereof, for example salts of the DNA or RNA. Derivatives for the purpose of the inventions are sequences or parts thereof which have been modified, for example, by methods of molecular biology and adapted to the particular requirements, for example truncated genes or parts of genes (for example promoter sequences, terminator sequences), cDNAs or chimeras thereof, constructs for expression and cloning and salts thereof.

One embodiment relates to the genomic sequences (genes) of the type L semaphorins. The invention relates to the intron and exon sequences and gene-regulatory sequences, for example promoter, enhancer and silencer sequences.

This embodiment relates on the one hand to the gene of H-SemaL or its derivatives. The invention relates on the one hand to a gene which comprises the nucleotide sequence given in Table 14. The invention further relates to the gene which comprises the nucleotide sequence which is deposited in the GenBank® databank under accession number AF030697.

This embodiment further relates to the gene of M-SemaL and its derivatives.

The invention further relates to the cDNA of H-SemaL or its derivatives (for example parts of the cDNA). A particular embodiment is the cDNA of H-SemaL according to the nucleotide sequence in Table 2. The invention further relates to the cDNA of H-SemaL which is deposited in the GenBank® databank under accession number AF030698. The invention also relates to the mRNAs corresponding to these cDNAs, or parts thereof.

The invention further relates to the cDNA of M-SemaL or its derivatives (for example parts of the cDNA). A particular embodiment is the partial cDNA sequence of M-SemaL shown in Table 3, and cDNA sequences which comprise this partial cDNA sequence. Another embodiment of the invention relates to the cDNA of M-SemaL which is deposited in the GenBank databank under accession number AF030699. The invention also relates to the mRNAs corresponding to these cDNAs, or parts thereof.

The invention also comprises alleles and/or individual expression forms of the genes/mRNAs/cDNAs which differ only slightly from the semaphorin sequences described herein and code for an identical or only slightly modified protein (difference in the amino acid sequence less than or equal to 10%) (further example of derivatives). Further examples of the derivatives are given

by the constructs indicated in the examples. The sequences of these constructs are depicted in Tables 7 to 14 and can be interpreted taking account of the annotation for plasmids.

The invention further relates to plasmids which comprise DNA which codes for the type L semaphorins or derivatives thereof. Plasmids of this type may be, for example, plasmids with high replication rates suitable for amplification of the DNA, for example in E. coli.

A specific embodiment comprises expression plasmids with which the semaphorins or parts thereof or their derivatives can be expressed in prokaryotic and/or eukaryotic expression systems. Both constitutive expression plasmids and those containing inducible promoters are suitable.

The invention also relates to processes for preparing nucleic acids which code for type L semaphorins or derivatives thereof.

These nucleic acids, for example DNA or RNA, can be synthesized, for example, by chemical means. In particular, it is possible for these nucleic acids, for example the corresponding genes or cDNAs or parts thereof, to be amplified by PCR using specific primers and suitable starting material as template. (For example cDNA from a suitable tissue or genomic DNA).

A specific process for preparing semaphorin L cDNA and the H-SemaL gene is described in the examples.

The invention also relates to processes for preparing type L semaphorins. For example, a semaphorin L or a derivative thereof can be prepared by cloning a corresponding nucleic acid sequence which codes for a type L semaphorin or a derivative thereof into an expression vector and using the latter recombinant vector to transform a suitable cell. It is possible to use, for example, prokaryotic or eukaryotic cells. The type L semaphorins or derivatives thereof may also, where appropriate, be prepared by chemical means.

In addition, the type L semaphorins and derivatives thereof can be expressed as fusion proteins, for example with proteins or peptides which permit detection of the expressed fusion protein, for example as fusion protein with GFP (green fluorescent protein). The semaphorins may also be expressed as fusion proteins with one, two, three or more epitope tags, for example with Myc and/or His (6xhistidine) and/or flu tags. It is correspondingly possible to use or prepare plasmids which comprise DNA sequences which code for these fusion proteins. For example, semaphorin-encoding sequences can be cloned into plasmids which contain DNA sequences which code for GFP and/or epitope tags, for example Myc tag, His tag, flu tag. Specific examples thereof are given by the examples and the sequences listed in the tables, where appropriate with the assistance of the annotation relating to the plasmids.

The invention further relates to antibodies which specifically bind or recognize the type L semaphorins, derivatives thereof or parts thereof. Possible examples thereof are polyclonal or monoclonal antibodies which can be produced, for example, in mouse, rabbit, goat, sheep, chicken etc.

A particular embodiment of this subject-matter of the invention comprises antibodies directed against the epitopes which correspond to the amino acid sequences from position 179 to 378 or 480 to 666 of the H-SemaL sequence shown in Table 4. The invention also relates to a process for preparing specific anti-semaphorin L antibodies, using for the preparation antigens comprising said epitopes.

The invention also relates to processes for preparing the antibodies, preferably using for this purpose a fusion protein consisting of a characteristic semaphorin epitope and an epitope tag which can be used for the subsequent purification of the recombinant fusion protein. The purified fusion protein can subsequently be used for the immunization. To prepare the recombinant fusion protein, a corresponding recombinant expression vector is prepared

and used to transform a suitable cell. The recombinant fusion protein can be isolated from this cell. The procedure can be, for example, like that described in Example 8.

These antibodies can be used, for example, for purifying the corresponding semaphorins, for example H-SemaL and its derivatives, for example on affinity columns, or for the immunological detection of the proteins, for example in an ELISA, in a Western blot and/or in immunohistochemistry. The antibodies can also be used to analyze the expression of H-SemaL, for example in various cell types or cell lines.

The cDNA of H-SemaL has a length of 2636 nucleotides (Table 2). The gene product of the H-SemaL cDNA has a length of about 666 amino acids (Table 4) and displays the typical domain structure of a type L semaphorin. The gene product has an N-terminal signal peptide (amino acids 1 to 44), Sema domain (amino acid 45 to approximately amino acid 545), and Ig (immunoglobulin) domain (approximately amino acids 550 to 620) and, at the C-terminal end, a hydrophobic amino acid sequence which represents a potential transmembrane domain. This domain structure has never previously been described for semaphorins. It relates to a membrane-associated glycoprotein which is probably located on the cell surface and belongs to a new subgroup. On the basis of this previously unknown domain structure, the semaphorins can now be divided into VI subgroups:

I		Secreted, without other domains (for example ORF-A49)
II	Ig	Secreted (without transmembrane domain) (for example AHV-Sema)
III	Ig, TM, CP	Membrane-anchored with cytoplasmic sequence (for example CD100)
IV	Ig, (P), HPC	Secreted with hydrophilic C terminus (for example H-Sema-III, M-SemaD, collapsin-1)
V	Ig, TM, CP	Membrane-anchored with C-terminal 7 thrombospondin motif (for example M-SemaF and G)

VI	Ig, TM	Membrane-anchored (for example H-SemaL, M-SemaL)

The unglycosylated, unprocessed form of H-SemaL has a calculated molecular weight of about 74.8 kd (74823 dalton) (calculated using Peptide-Sort, GCG program package). The isoelectric point is calculated to be pH = 7.56.

A possible signal peptide cleavage site is located between amino acids 44 and 45 (Table 3; calculated with SignalP (http.//www.cbs.dtu.dk/services/Signal P), a program based on neural networks for analyzing signal sequences {Nielsen H. et. al. (1997) Protein Engineering 10:1-6}). This gives for the processed protein (without signal peptide) a molecular weight (MW) of 70.3 kd (70323 dalton) and an isoelectric point of pH=7.01.

The genomic structure is likewise substantially elucidated. The H-SemaL gene has 13 or 15 or more exons, preferably 14 exons, and 12 or 14 introns, preferably 13 introns. Because of this complex exon-intron structure, various splicing variants are possible. The mRNA of the transcribed H-SemaL gene is found in the Northern blot particularly in placenta, gonads, thymus and spleen. No mRNA has been detected in neuronal tissue or in muscle tissue. There is evidence of specifically regulated expression in endothelial cells.

Alternative splicing may also result in forms of H-SemaL with intracytoplasmic sequences which are involved in intracellular signal transduction, similar to, for example, CD100. It would likewise be possible for alternative splicing to result in secreted forms of H-SemaL, analogous to viral AHV-Sema.

Nucleotide and amino acid sequence analyses were performed with the aid of the GCG program package (Genetics Computer Group (1991) Program manual for the GCG package, Version 7, 575 Science Drive, Wisconsin, USA 53711), FASTA (Pearson and Lipman (1988) Proc. Natl. Acad. Sci. 85, 2444-

2448) and BLAST program (Gish and States (1993) Nat. Genet.3, 266-272; Altschul et al. (1990) J. Mol. Biol. 215, 403-410). These programs were also used for sequence comparisons with GenBank (Version 102.0) and Swiss Prot (Version 34.0).

Post-translational modifications such as glycosylation and myristylation of H-SemaL are likewise possible. Consensus sequences for N-glycosylation sites were found with the aid of the Prosite program (GCG program package) at positions 105, 157, 258, 330 and 602 of the amino acid sequence of H-SemaL (shown in Table 4), and those for myristylation were found at positions 114, 139, 271, 498, 499, 502 and 654 (consensus sequence: G~(E, D, R, K, H, P, F, Y, W) x (S, T, A,G, C, N)~(P)). In addition, the amino acid sequence of H-SemaL contains several consensus sequences for potential phosphorylation sites for various kinases. It can therefore be assumed that H-SemaL can be the substrate of various kinases, for example phosphorylation sites for creatine kinase 2, protein kinase C and tyrosine kinase.

Predicted creatine kinase 2 phosphorylation sites (consensus sequence Ck2: (S,T)x2(D,E)) (Prosite, GCG) at positions 119, 131, 173, 338, 419 and 481 of the amino acid sequence.

Predicted protein kinase C phosphorylation sites (consensus sequence PkC: (S,T)x(R,K)) (Prosite, GCG) at positions 107, 115, 190, 296, 350, 431, 524 and 576 of the amino acid sequence.

Predicted tyrosine kinase phosphorylation site (consensus sequence: (R,K)x{2,3}(D,E)x{2,3}Y) (Prosite, GCG) at position 205 of the amino acid sequence.

The consensus sequences are indicated in the single letter code for amino acids.

An "RGD" motif (arginine-glycine-aspartic acid) characteristic of integrins is located at position 267.

The glycosylation sites are highly conserved between viral AHV-Sema, H-SemaL and (as far as is known) M-SemaL.

Di- or multimerization of H-SemaL is possible and has been described for other semaphorins such as CD100 {Hall et al. (1996)}. The CD100 molecule is likewise a membrane-anchored glycoprotein dimer of 150kd. However, CD100 is not closely related to the human semaphorin (H-SemaL) according to the invention.

The partial cDNA sequence of M-SemaL has a length of 1195 nucleotides. This sequence codes for a protein having 394 amino acids. These 394 amino acids correspond to amino acids 1 to 396 of H-SemaL. The signal peptide in M-SemaL extends over amino acids 1 to 44 (exactly as in H-SemaL). The Sema domain starts at amino acid 45 and extends up to the end or probably beyond the end of the sequence shown in Table 4.

Multiple alignments were carried out using the Clustal W program (Thompson et al. (1994)). These alignments were processed further manually using SEAVIEW (Galtier et al. (1996) Comput. Appl. Biosci 12, 543-548). The phylogenetic distances were determined using Clustal W (Thompson et al. (1994)).

Comparison of the protein sequences of the known and of the novel semaphorins and phylogenetic analysis of these sequences shows that the genes can be categorized according to their phylogenetic relationship. The C-terminal domain structure of the corresponding semaphorin subtypes is, of course, involved in this as a factor deciding why semaphorins in the same subgroups are, as a rule, also more closely related phylogenetically than are semaphorins in different subgroups. The species from which the semaphorin

was isolated also has an influence, i.e. whether the corresponding species are phylogenetically closely related to one another or not.

A phylogenetic analysis (compare Figure 3) of the known semaphorin amino acid sequences (complete sequences and/or part-sequences, using the amino acid sequences for H-SemaL and M-SemaL shown in Tables 4 and 5 and for all other sequences the sequences stored under the accession numbers or the encoded amino acid sequences derived from these sequences) using the CLUSTAL W program {Thompson J.D. et al. (1994) Nucleic Acids Res. 22:4673-4680} shows that the amino acid sequences of H-SemaL and M-SemaL are phylogenetically closely related to one another and form a separate phylogenetic group. H-SemaL and M-SemaL in turn are phylogenetically most closely related to AHV-Sema and Vac-A39. The are distinctly more closely related to one another than to any other previously disclosed semaphorin. The analysis also shows that other semaphorins are also phylogenetically closely related to one another and form separate groups within the semaphorins. For example, the semaphorins which are secreted, for example H-Sema III, -IV, -V and -E belong in one phylogenetic group. Their homologs in other species also belong to this subfamily, whereas the human (transmembrane) CD100 belongs in one phylogenetic group together with the corresponding mouse homolog (M-SemaG2) and with Collapsin-4.

In relation to the complete amino acid sequences, the observed homologies within the phylogenetic groups are between about 90% and 80% amino acid identity in relation to very closely related genes such as, for example, H- and M-SemaE or -III/D and somewhat less than 40% in the case of less related genes of the semaphorins. Within the Sema domain, the observed amino acid identity is a few percent higher, and, owing to its great contribution to the total protein (50-80% of the protein belong to the Sema domain) of the amino acid sequence, this considerably influences the overall identity.

H-SemaL is, calculated for the complete protein, 46% identical with AHV-Sema, but if the Sema domain is considered on its own, then the amino

acid identity is 53%. This is higher than, for example, between the related M-Sema-B and -C (37% identity in relation to the complete protein, 43% identity in relation to the Sema domain), similar to M-SemaA and -E (43% complete protein, 53% Sema domain). The amino acid identity between the partial M-SemaL sequence (Table 6) and H-SemaL (Table 5) in the region of the Sema domain is 93% so that it can be assumed that the correspondingly homologous mouse gene is involved.

Semaphorins corresponding to H-SemaL and M-SemaL in other species may have an amino acid identity within the Sema domain of more than 40% in relation to H-SemaL. In closely related vertebrates (mammals, birds) amino acid identities above 70% may even be found.

The semaphorins belong to a new subfamily with greater amino acid identity to the viral AHV-Sema than to the previously disclosed human and murine semaphorins, and with a C-terminal structure not previously disclosed for human semaphorins. These novel semaphorins (members of the subfamily) are distinguished by belonging, because of their domain structure, to subgroup IV and/or to the same phylogenetic group as H-SemaL and M-SemaL and/or have, in relation to the complete amino acid sequence, an amino acid identity of at least 30 to 40%, preferably 50 to 60%, particularly preferably 70 to 80%, or a greater identity, to H-SemaL and/or have, in relation to the Sema domain, an amino acid identity of at least 70%, preferably greater than 80%, particularly preferably greater than 90%, to H-SemaL.

The type L semaphorins also have a different type of biochemical function. One novel function of these semaphorins is modulation of the immune system.

The closest relative of H-SemaL is the viral AHV semaphorin (AHV-Sema). The latter has a similar size but, in contrast to H-SemaL, has no transmembrane domain. AHV-Sema is presumably secreted by virus-infected

cells in order to block the H-SemaL equivalent receptor (type L semaphorin in the blue wildebeest) in the natural host (blue wildebeest) and thus elude the attack of the immune system. It is also conceivable that there is a function as repulsive agent (chemorepellant) for cells of the immune system.

The biochemical function of the novel type L semaphorins and derivatives thereof is to be regarded as generally immunomodulating and/or inflammation-modulating. They are able on the one hand

A) as molecules inhibiting the immune response to display their effect as chemorepellant and/or immunosuppressant either locally, for example as transmembrane protein on the surface of cells, or else over larger distances, for example if they are secreted due to processing (for example proteases) or alternative splicing, for example by diffusion in the tissue.

For example, expression of these novel type L semaphorins for example on the surface of the cells of the vascular endothelium can prevent leukocyte attachment and migration thereof through the vessel wall. The novel semaphorins may play a part in maintenance of barrier effects, for example to prevent infections in particularly "important" or exposed organs, for example to maintain the blood-brain barrier, the placental circulation and/or other immunologically privileged locations (for example pancreatic islets) and/or in prevention of autoimmune diseases. In addition, the novel semaphorins and/or their derivatives may also be involved in repulsive signals in various tissues, for example for cells of the immune system (for example leukocytes) to prevent inadvertent activation of defense mechanisms.

B) In addition, the novel semaphorins and/or derivatives thereof may have functions as accessory molecules. Expressed on the cell surface, they may, for example, be involved in the interaction with cells of the

immune system as part of the activation of defense mechanisms, for example in cases of virus infection.

This reveals several possible uses of the novel type L semaphorins and derivatives thereof, and the nucleic acids coding for these proteins.

Function A): This comprises an immunosuppressant and/or anti-inflammatory principle: there are numerous potential possibilities of use in the areas of organ transplantation, therapy of inflammations, immunotherapy and gene therapy.

For example, nonhuman, transgenic animals can be produced with the aid of the semaphorin-encoding DNA or derivatives thereof.

One possible use of these animals is in the inhibition of transplant rejection in transgenic models of organ transplantations. For example, transgenic animal organs protected against rejection can be produced for xenotransplantations. This ought to be possible for example also together with other transgenes (for example complement regulators such as DAF or CD59). Another use is in the production of nonhuman knock-out animals, for example knock-out mice ("Laboratory Protocols for Gene-Targeting", Torres and Kühn (1997) Oxford University Press, ISBN 0-19-963677-X): It is possible by knocking out the mouse M-SemaL gene for example to find other functions of the gene. They also represent potential model systems for inflammatory diseases if the mice can survive without semaphorin gene. If M-SemaL is important for immunomodulation, a plurality of such mice is to be expected. In addition, nonhuman knock-in animals, for example mice, can be produced. This entails, for example, replacing M-SemaL by normal/modified H-SemaL or modified M-SemaL (for example integration of the novel semaphorin subtypes under the control of constitutive and/or inducible promoters). Animals of this type can be used, for example, for looking for further functions of the novel semaphorins, for example functions of the human gene or derivatives of these genes, or be used for identifying and characterizing immunomodulating agents.

Use of, for example, nucleic acids which code for type L semaphorins or derivatives thereof for producing, for example, recombinant immunosuppressants, other soluble proteins or peptides derived from the amino acid sequence of type L semaphorins, for example from H-SemaL or the corresponding nucleic acids, for example genes. It is also possible in a similar way to produce agonists with structural similarity. These immunosuppressant agents or agonists may be used for autoimmune diseases and inflammatory disorders and/or organ transplantations too.

Gene therapy with type L semaphorins, for example with nucleic acids which code for H-SemaL or derivatives thereof, for example using viral or nonviral methods. Use in autoimmune diseases and inflammatory disorders, the transduction of organs and before/during/after transplantations to prevent transplant rejection.

It is particularly possible to employ the novel semaphorins and/or the nucleic acids coding for these semaphorins, and derivatives thereof, in particular H-SemaL, DNA coding for H-SemaL, and derivatives thereof, in a method for screening for agents, in particular for identifying and characterizing immunomodulating agents.

Function B): H-SemaL is an accessory molecule which is expressed on the cell surface and is involved in the interaction with cells, for example of the immune system, for example as accessory molecule in the activation of signal pathways. A viral gene or the gene product of a viral or other pathogenic gene, for example of microbiological origin, might act, for example, as competitive inhibitor of this accessory molecule. One use of the novel semaphorins with this function is likewise in the area of organ transplantation, therapy of inflammation, immunotherapy and/or gene therapy.

For example, the novel semaphorins can be used in a method for screening for antagonistic agents or inhibitors. Agents identified in this way can then be

employed, for example, for blocking the semaphorin receptor. Soluble and/or secreted H-SemaL antagonists or inhibitors may be, for example, chemical substances or the novel semaphorins or derivatives thereof themselves (for example parts/truncated forms thereof, for example without membrane domain or as Ig fusion proteins or peptides derived from the latter, which are suitable for blocking the corresponding receptor). Specific antagonists and/or inhibitors identified in this way may, for example, have competitive effects and be employed for inhibiting rejection, for example in transgenic models of organ transplantations and for autoimmune diseases, inflammatory disorders and organ transplantations. Nucleic acids, for example DNA, which code for the novel semaphorins, or derivatives thereof produced with the aid of methods of molecular biology, may be used, for example, for producing nonhuman transgenic animals. Overexpression of H-SemaL in these transgenic animals may lead to increased susceptibility to autoimmune diseases and/or inflammatory disorders. Such transgenic animals are thus suitable for screening for novel specific immunomodulating agents.

Such nucleic acids can likewise be used to produce nonhuman knock-out animals, for example knock-out mice in which the mouse M-SemaL gene is switched off. Such knock-out animals can be employed to search for further biochemical functions of the gene. They also represent potential model systems for inflammatory disorders if the mice are able to survive without the M-SemaL gene.

This DNA can likewise be used to produce nonhuman knock-in animals, for example mice. This entails the M-SemaL gene being replaced by a modified M-SemaL gene/cDNA or an optionally modified, for example mutated, type L semaphorin gene/cDNA of another species, for example H-SemaL. Such transgenic animals can be used to look for further functions of the semaphorins according to the invention.

The invention also relates to the use of the type L semaphorins and derivatives thereof, and of the nucleic acids coding for these proteins, for

example genes/cDNAs and derivatives thereof and/or agents identified with the aid of these semaphorins for producing pharmaceuticals. It is possible, for example, to produce pharmaceuticals which can be used in gene therapy and which comprise agonists and/or antagonists of the expression of the type L semaphorins, for example of H-SemaL. It is possible to use for this purpose, for example, viral and/or nonviral methods. These pharmaceuticals can be employed, for example, for autoimmune diseases and inflammatory disorders, organ transplantations before and/or during and/or after the transplantation to prevent rejection.

The nucleic acids coding for the novel semaphorins, for example genes, cDNAs and derivatives thereof, can also be employed as aids in molecular biology.

In addition, the novel semaphorins, especially H-SemaL and nucleic acids, for example genes/cDNAs thereof can be employed in methods for screening for novel agents. Modified proteins and/or peptides derived, for example, from H-SemaL and/or M-SemaL can be used to look for the corresponding receptor and/or its antagonists or agonist in functional assays, for example using expression constructs of H-SemaL and homologs.

The invention also relates to the use of a type L semaphorin or a nucleic acid sequence which codes for a type L semaphorin in a method for identifying pharmacological agents, especially immunomodulating agents.

The invention also relates to methods for identifying agents employing a type L semaphorin or a derivative thereof or a nucleic acid sequence which codes for a type L semaphorin, or a derivative thereof, in order to identify pharmacological agents, for example immunomodulating agents. The invention relates, for example, to a method in which a type L semaphorin is incubated under defined conditions with an agent to be investigated and, in parallel, a second batch is carried out without the agent to be investigated but

under conditions which are otherwise the same, and then the inhibiting or activating effect of the agent to be investigated is determined.

The invention also relates, for example, to methods for identifying agents where a nucleic acid sequence which codes for a type L semaphorin or a derivative thereof is expressed under defined conditions in the presence of an agent to be investigated, and the extent of the expression is determined. It is also possible, where appropriate, in such a method to carry out two or more batches in parallel under the same conditions but with the batches containing different amounts of the agent to be investigated.

For example, the agent to be investigated may inhibit or activate transcription and/or translation.

The type L semaphorin can, like its viral homologs, bind to the newly described receptor molecule VESPR (Comeau et al, (1998) Immunity, Vol. 8, 473-482) and in monocytes can presumably cause induction of cell adhesion molecules such as ICAM-1 and cytokines such as interleukin-6 and interleukin-8. This may lead to activation thereof and to cell aggregation. The expression pattern of the VESPR receptor shows some interesting parallels with H-SemaL, for example strong expression in placenta and pronounced expression in spleen tissue. Interactions with other as yet unknown receptors of the plexin family or other receptors are possible. It may also interact with itself or other semaphorin-like molecules. Interaction of the type L semaphorins may take place in particular via a conserved domain in the C-terminal region of the Sema domain.

Concerning the annotation on plasmids:

pMelBacA-H-SemaL (6622bp) in pMelBacA (Invitrogen, De Schelp, NL) (SEQ ID NO.42). Nucleotide 96-98 ATG – start codon, nucleotide 96-168 mellitin signal sequence, nucleotide 168-173 BamHI cleavage site (PCR/cloning), nucleotide 171-1998 reading frame SEMA-L amino acids 42-649 (without own

signal sequence and without transmembrane sequence), nucleotide 1993-1998 EcoRI cleavage site (PCR/cloning) and nucleotide 1992-1994 stop codon

Plasmid pCDNA3.1-H-SemaL-MychisA (7475 bp) (SEQ ID NO. 35): nucleotide 954-959 BamHI cleavage site (cloning), nucleotide 968-970 ATG SEMAL, nucleotide 968-2965 reading frame SEMAL, nucleotide 2963-2968 Pml I cleavage site, nucleotide 2969-2974 HindIII cleavage site, nucleotide 2981-3013 Myc tag, nucleotide 3026-3033 6xHis tag, nucleotide 3034-3036 stop codon,

Plasmid pCDNA3.1-H-SemaL-EGFP-MychisA (8192 bp):(SEQ ID NO. 36): nucleotide 954-959 BamHI cleavage site (cloning), nucleotide 968-970 ATG SEMA-L, nucleotide 968-2965 reading frame SEMA-L, nucleotide 2963-2965 half Pml I cleavage site, nucleotide 2966-3682 reading frame EGFP (cloned in Pml I), nucleotide 3683-3685 half Pml I cleavage site, nucleotide 3685-3691 HindIII, nucleotide 3698-3730 Myc tag, nucleotide 3743-3760 6xHis tag, and nucleotide 3761-3763 stop codon

Plasmid pIND-H-SemaL-EA (7108 bp) in vector pIND (Invitrogen, De Schelp, NL) (SEQ ID No. 38): nucleotide 533-538 BamHI cleavage site (cloning), nucleotide 546-548 ATG SEMA-L, nucleotide 546- reading frame SEMA-L, nucleotide 2542-2547 Pml I cleavage site, nucleotide 2548-2553 HindIII cleavage site and nucleotide 2563-2565 stop codon.

Plasmid pIND-H-SemaL-EE (total length 7102 bp) in vector pIND (Invitrogen, De Schelp, NL) (SEQ ID No. 37): nucleotide 533-538 BamHI cleavage site (cloning), nucleotide 546-548 ATG SEMA-L, nucleotide 546- reading frame SEMA-L, nucleotide 2542-2547 Pml I cleavage site, nucleotide 2548-2553 HindIII cleavage site, nucleotide 2560-2592 Myc tag, nucleotide 2605-2622 6xHis tag and nucleotide 2623-2625 stop codon.

Plasmid pQE30-H-SemaL-179-378.seq (4019 bp) in vector pQE30 (Qiagen, Hilden) corresponds to pQE30-H-SemaLBH (SEQ ID No. 39): nucleotide 115-117 ATG, nucleotide 127-144 6xHis tag, nucleotide 145-750 BamHI-HindIII PCR fragment SEMA-L amino acids (aa) 179-378 and nucleotide 758-760 stop codon.

Plasmid pQE31-H-SemaL- (SH (3999 bp) in vector pQE31 (Qiagen, Hilden) (SEQ ID No. 40): nucleotide 115-117 ATG, nucleotide 127-144 6xHis tag, nucleotide (147-152 BamHI), nucleotide 159-729 SacI-HindIII fragment SEMA-L (C-terminal) aa480-666 and nucleotide 734-736 stop codon.

Examples:

Experimental conditions used in the examples:

PCR programs used:

Taq52-60 (with Ampli-Taq^{R} polymerase, Perkin Elmer, Weil der Stadt, Germany)

96°C/60s	1 cycle
96°C/15s-52°C/20s-70°C/60s	40 cycles
70°C/60s	1 cycle

Taq60-30

96°C/60s	1 cycle
96°C/15s-60°C/20s-70°C/30s	35 cycles
70°C/60s	1 cycle

Taq60-60

96°C/60s	1 cycle
96°C/15s-60°C/20s-70°C/60s	35 cycles
70°C/60s	1 cycle

Taq62-40

96°C/60s	1 cycle
96°C/15s-62°C/20s-70°C/40s	35 cycles
70°C/60s	1 cycle

Reaction conditions used for PCR with Taq polymerase:

50µl reaction mixtures with 100-200ng of template, 200µM dNTP, 0.2-0.4 µM each primer, 2.5U of Ampli-Taq^{R}, 5µl of the 10x reaction buffer supplied

Programs used for:

1. XL62-6 (with expand-long template PCR System^{R}, Boehringer Mannheim, Germany)

94°C/60s	1 cycle
94°C/15s-62°C/30s-68°C/6min	10 cycles
94°C/15s-62°C/30s-68°C/(6min+15s/cycle)	25 cycles
68°C / 7min	1 cycle

2. XL62-12 (with expand-long template PCR System^{R}, Boehringer Mannheim, Germany)

94°C/60s	1 cycle
94°C/15s-62°C/30s-68°C/12min	10 cycles
94°C/15s-62°C/30s-68°C/(12min+15s/cycle)	25 cycles
68°C / 7min	1 cycle

Reaction conditions for PCR with expand-long template PCR System
50µl reaction mixtures with 100-200ng of template, 500µM dNTP, 0.2-0.4 µM each primer, 0.75µl of enzyme mix, 5µl of the 10x reaction buffer No. 2 supplied.

Example 1:

Starting from AHV-Sema sequences (Ensser & Fleckenstein (1995), J. General Virol. 76: 1063-1067), PCRs and RACE-PCRs were carried out. The starting material used for this was human cDNA from placental tissue onto which adaptors had been ligated for the RACE amplification (Marathon^{TM}-cDNA Amplification Kit, Clontech Laboratories GmbH, Tullastraße 4, 69126 Heidelberg, Germany). Firstly specific primers (No. 121234 + No. 121236, Table 6) were used to amplify a PCR fragment with a length of about 800bp (base pairs) (PCR program: (Taq60-60)). This was cloned and sequenced (Taq dye-deoxy terminator sequencing kit, Applied Biosystems, Foster City, CA, USA/ Brunnenweg 13, Weil der Stadt). Sequencing of the PCR product revealed a sequence which has a high degree of homology with the DNA sequence of AHV-Sema, identical to the sequence of the two ESTs.

A PCR fragment of 600bp was identified using the primer pair (No. 121237 + No. 121239, Table 6). It emerged that they were clones with DNA sequences from the same gene.

Example 2:

The 800bp PCR fragment from Example 1 was radiolabeled (random priming by the method of {Feinberg (1983) Anal. Biochem. 132:6-13}, with ^{32}P-α-dCTP) and used as probe for a multitissue Northern blot (Human Multiple Tissue Northern Blot II, Clontech, Heidelberg, Germany) which contains mRNA samples from the tissues spleen, thymus, prostate, testes, ovaries, small intestine, large intestine and leukocytes (PBL). This clearly showed expression of an mRNA with a length of about 3.3kb in spleen and gonads (testes, ovaries), and less strongly in the thymus and intestine. Hybridization of a master blot (dot-blot with RNA from numerous tissues (Human RNA Master Blot™, Clontech)) confirmed this result and also showed strong expression in placental tissue.

Hybridization was carried out under stringent conditions (5xSSC, 50 mM Na phosphate pH 6.8, 50% formamide, 100 µg/ml yeast RNA) at 42°C for 16 hours. The blots were washed stringently (65°C, 0.2XSSC, 0.1% SDS) and exposed to a Fuji BAS2000 Phosphoimager™.

Example 3:

A cDNA library from human spleen, cloned in the bacteriophage Lambda gt10 (Human Spleen 5' STRETCH PLUS cDNA, Clontech), was screened with this probe, and a lambda clone was identified. The cDNA with a length of 1.6kb inserted in this clone was amplified by PCR (Expand™ Long Template PCR System, Boehringer Mannheim GmbH, Sandhofer Straße 116, 68305 Mannheim) using the vector-specific primers No. 207608 + No. 207609 (Table 6) (flanking the EcoRI cloning site), and the resulting PCR fragment was sequenced. This clone contained the 5' end of the cDNA and also extended

the known cDNA sequence in the 3' direction. Starting from the new part-sequences of the cDNA, new primers for the RACE-PCR were developed (No. 232643, No. 232644, No. 233084, Table 6). Together with an improved thermocycler technique (PTC-200 from MJ-Research, Biozym Diagnostik GmbH, 31833 Hess. Oldendorf) with distinctly better performance data (heating and cooling rates), a 3' RACE-PCR product was amplified using the primers No. 232644 and No. 232643 and AP1, and was cloned into the vector pCR2.1 (Invitrogen, De Schelp 12, 9351 NV Leek, The Netherlands). The 3' RACE-PCR product was sequenced and the 3' end of the cDNA was identified in this way. A RACE amplification in the 5' direction (primers No. 131990 and No. 233084 and AP1) extended the 5' end of the cDNA by a few nucleotides and confirmed the amino terminus of H-SemaL found in the identified lambda clone.

Example 4:

Starting from a short murine EST (Accession No. AA260340) and a primer derived therefrom, No. 260813 (Table 6) and the H-SemaL specific primer No. 121234 (Table 6), PCR (conditions: Taq52-60) was used to amplify a DNA fragment with a length of about 840 bp of murine cDNA, followed by cloning into the vector pCR2.1. The gene containing this DNA fragment was called M-SemaL. The resulting M-SemaL DNA fragment was used to investigate a cDNA bank from mouse spleen (Mouse Spleen 5' STRETCH cDNA, Clontech), identification of several clones being possible.

PCR (Taq60-30) with the primers No. 260812 and No. 260813 from murine endothelial cDNA provided a PCR fragment with a length of 244 base pairs. The PCR results showed that there is distinct baseline expression in murine endothelial cells which declines after stimulation with the cytokine interferon-γ and lipopolysaccharides.

Example 5:

Investigations on the location in the chromosome were carried out by fluorescence in situ hybridization (FISH). For this purpose, human and murine metaphase chromosomes were prepared starting from a human blood sample and the mouse cell line BINE 4.8 (Keyna et al. (1995) J. Immunol. 155, 5536-5542), respectively (Kraus et al. (1994) Genomics 23, 272-274). The slides were treated with RNase and pepsin (Liehr et al. (1995) Appl. Cytogenetics 21, 185-188). For the hybridization, 120 mg of human nick-translated semaphorin sample and 200 mg of a corresponding mouse sample were used. The hybridization was in each case carried out in the presence of 4.0 µg of COT1-DNA and 20 µg of STD at 37°C (3 days) in a moistened chamber.

The slides were washed with 50% formamide/2x SSC (3 times for 5 min each time at 45°C) and then with 2x SSC (3 times for 5 min each time at 37°C), and the biotinylated sample was detected using the FITC-avidin system (Liehr et al. (1995)). The slides were evaluated using a fluorescence microscope. 25 metaphases/sample were evaluated, carrying out each experiment in duplicate. It emerged that H-SemaL is located on chromosome 15q23. Located adjacent in the chromosome is the locus for Bardet-Biedls syndrome and Tay-Sachs disease (hexosaminidase A).

Example 6:

The genomic intron-exon structure of the H-SemaL gene is for the most part elucidated.

Genomic DNA fragments were amplified starting from 250 mg of human genomic DNA which had been isolated from PHA-stimulated peripheral lymphocytes (blood). Shorter fragments were amplified using Ampli Taq^{R} (Perkin Elmer), and longer fragments were amplified using the expanded long template PCR System^{R} (Boehringer Mannheim).

It has been possible by PCR amplification to date to clone and characterize almost the complete genomic locus of H-SemaL. It has already been possible in total to determine more than 8888 bp of the genomic sequence and thus substantially to elucidate the intron-exon structure of the gene.

Example 7:

Expression clonings:
Since no complete clone of the semaphorin gene could be isolated from the lambda-gt10 cDNA bank, and no complete clone was obtainable by PCR either, the coding region of the cDNA was amplified in 2 overlapping subfragments by PCR (XL62-6) using the primers No. 240655 and No. 121339 for the N-terminal DNA fragment, and the primers No. 240656 (contains HindIII and PmeI cleavage sites) and No. 121234 for the C-terminal DNA fragment. The resulting DNA fragments (subfragments) were cloned into the vector pCR21. The two subfragments were completely sequenced and finally the complete H-SemaL cDNA was prepared by inserting a 0.6kb C-terminal SstI-HindIII restriction fragment into the plasmid which contained the N-terminal DNA fragment and had been cut with the restriction enzymes SstI and HindIII. From this plasmid pCR2.1-H-SemaL (sequence shown in Table 7, SEQ ID NO. 34), the complete gene was cut out using the EcoRI cleavage site (in pCR2.1) and HindIII cleavage site (in primer No. 240656, Table 6) and ligated into a correspondingly cut constitutive expression vector pCDNA3.1(-)MycHisA (Invitrogen). The EcoRI-ApaI fragment (without Myc-His tag) was cut out of the resulting recombinant plasmid pCDNA3.1(-)H-SemaL-MycHisA (sequence shown in Table 8) and ligated into the inducible vector pIND (Ecdysone-Inducible Mammalian Expression System, Invitrogen) which had previously likewise been cut with EcoRI-ApaI. The recombinant plasmid was called pIND-H-SemaLEA (sequence shown in Table 11). An EcoRI-PmeI fragment (with Myc-His tag) from pCDNA3.1(-)H-SemaL-Myc-HisA (sequence shown in Table 9) was inserted into an EcoRI-EcoRV-cut vector pIND. The recombinant plasmid was called pIND-H-SemaL-EE (sequence shown in Table 10).

A fusion gene of H-SemaL with enhanced green fluorescent protein (EGFP) was prepared by ligating the PCR-amplified EGFP reading frame (from the vector pEGFP-C1 (Clontech), using the primers No. 243068 + No. 243069, Taq52-60) into the PmeI cleavage site of the plasmid pCDNA3.1(-)H-SemaL-MycHisA, resulting in the plasmid pCDNA3.1(-)H-SemaL-EGFP-MycHisA (sequence shown in Table 9).

Small letters in Tables 7 to 13 and Table 15 denote the sequence of H-SemaL, parts or derivatives thereof, and large letters denote the sequence of the plasmid.

Example 8:

To prepare H-SemaL-specific antibodies, cDNA fragments of H-SemaL were integrated into prokaryotic expression vectors and expressed in E. coli, and the semaphorin derivatives were purified. The semaphorin derivatives were expressed as fusion proteins with a His tag. Accordingly, vectors containing the sequence for a His tag and permitting integration of the semaphorin cDNA fragment into the reading frame were used. An N-terminal 6xhistidine tag makes it possible, for example, to purify by nickel chelate affinity chromatography (Qiagen GmbH, Max-Volmer Straße 4, 40724 Hilden):

1. The part of the H-SemaL cDNA coding for amino acids 179-378 was amplified by PCR using the primers No. 150788 and No. 150789, and this DNA fragment was ligated into the vector pQE30 (Qiagen) which had previously been cut with the restriction enzymes BamHI and HindIII (construct pQE30-H-SemaL-BH (sequence shown in Table 12)).

2. The section of the H-SemaL cDNA coding for the C-terminal amino acids 480-666 was cut with the restriction enzymes SstI and HindIII out of the plasmid pCR 2.1 and ligated into the vector pQE31 (Qiagen)

which had previously been cut with SstI and HindIII (construct pQE31-H-SemaL-SH (sequence shown in Table 13)).

Correct integration of the sequences in the correct reading frame was checked by DNA sequencing. The fusion proteins consisting of an N-terminal 6xhistidine tag and a part of the semaphorin H-SemaL were purified by Ni^{2+} affinity chromatography. The purified fusion proteins were used to immunize various animals (rabbit, chicken, mouse).

Example 9:

FACS analysis of various cell types (Figures 4 and 5)
The cells (about 0.2-0.5 x 10^6) were washed with FACS buffer (phosphate-buffered saline (PBS) with 5% fetal calf serum (FCS) and 0.1% Na azide) and then incubated with the antisera (on ice) for 1 hour in each case.

The primary antibodies used for the control (overlay chicken preimmune serum (1:50)) and for the specific detection (specific staining) comprised an H-SemaL-specific chicken antiserum (1:50). The specific antiserum with antibodies against amino acids (Aa) 179-378 (with N-terminal His tag) of H-SemaL was generated by immunizing chickens with the protein purified by Ni chelate affinity chromatography (as described in Example 8). The second antibody used was an FITC-labeled anti-chicken F(ab') antibody from rabbits (Dianova Jackson Laboratories, Order No. 303-095-006, Hamburg, Germany) (1 mg/ml). A rabbit anti-mouse IgG, FITC-labeled, was used for the CD100 staining. The second antibody was employed in each case in 1:50 dilution in FACS buffer.

The cells were then washed, resuspended in PBS and analyzed in the FACS. The FACS analysis was carried out using a FACS-track instrument (Becton-Dickinson). Principle: a single cell suspension is passed through a measuring channel where the cells are irradiated with laser light of 488 nm and thus fluorescent dyes (FITC) are excited. The measurements are of the light

scattered forward (forward scatter, FSC: correlates with the cell size), and to the side (sideward scatter, SSC: correlates with the granular content: different in different cell types) and fluorescence in channel 1 (FL 1) (for wavelengths in the FITC emission range, max. at 530 nm). 10,000 events (cells) were measured in this way each time.

The dot plot (Figures 4a-k) (figure on the left in each case): FSC against SSC (size against granular content/scatter) with, inside the boundary, the (uniform) cell population of similar size and granular content analyzed in the right-hand window (relevant right-hand figure in each case). The right-hand window shows the intensity of FL 1 (X axis) against the number of events (Y axis), that is to say a frequency distribution.

In each of these, the result with the control serum (unfilled curve) is superimposed on the result of the specific staining (filled curve). A shift of the curve for the specific staining to the right compared with the control corresponds to an expression of H-SemaL in the corresponding cells. A larger shift means stronger expression.

Cell lines used for FACS analysis:

a) U937 cell line
American Type Culture Collection ATCC; ATCC number: CRL-1593
Name: U-937
Tissue: lymphoma; histiocytic; monocyte-like
Species: human;
Depositor: H. Koren

b) THP-1 cell line
ATCC number: TIB-202
Tissue: monocyte; acute monocytic leukemia
Species: human
Depositor: S. Tsuchiya

c) K-562 cell line
ATCC number: CCL-243
Tissue: chronic myelogenous leukemia
Species: human;
Depositor: H.T. Holden

d) L-428 cell line
DSMZ-Deutsche Sammlung von Mikroorganismen und Zellkulturen GmbH,
DSMZ No: ACC 197
Cell type: human Hodgkin's lymphoma

e) Jurkat cell line
DSMZ-Deutsche Sammlung von Mikroorganismen und zellkulturen GmH,
DSMZ No: ACC 282
Cell type: human T cell leukemia

f) Daudi cell line
ATCC number: CCL-213
Tissue: Burkitt's lymphoma; B lymphoblast; B cells
Species : human
Depositor: G. Klein

g) LCL cell line
EBV-transformed lymphoblastoid B-cell line.

h) Jiyoye (P-2003) cell line
ATCC number: CCL-87
Tissue: Burkitt's lymphoma; B cells, B lymphocyte
Species: human
Depositor: W. Henle

i) CBL-Mix57

Human T-cell line (isolated from blood) transformed with recombinant H. Saimiri (wild-type without deletion)

j) CBL-Mix59
Human T-cell line (isolated from blood) transformed with H. Saimiri (deletion of ORF71).

Example 10: Protein gel and Western blot

Secretable human SEMA-L (amino acids 42-649 in Table 4 (without signal peptide and without transmembrane domain)) was cloned into the plasmid pMelBac-A (Invitrogen, De Schelp, Leck, The Netherlands, Cv 1950-20) and, in this way, the plasmid pMelBacA-H-SemaL (length 6622bp) was generated (Figure 8). The H-SemaL derivative was expressed in the baculovirus system (Bac-N-Blue, Invitrogen). Expression was carried out in the cell lines derived from insect egg cells Sf9 (from Spodoptera frugiperda) and High Five™ (from Trichoplusia ni, U.S. Pat. No. 5,300,435, purchased from Invitrogen) by infection with the recombinant, plaque-purified baculoviruses.

The expression was carried out in accordance with the manufacturer's instructions.

The proteins were then fractionated in a gel, and the H-SemaL derivative was detected in a Western blot. Detection was carried out with H-SemaL-specific chicken antiserum (compare Example 8 and Figure 7) (dilution 1:100). The specific chicken antibody was detected using anti-IgY-HRP conjugate (dilution: 1:3000, from donkey; Dianova Jackson Laboratories) in accordance with the manufacturer's instructions.

Example 11: Preparation of pMelBacA-H-SEMAL

The recombinant vector (pMelBacA-H-SEMAL, 6622bp) was prepared by cloning an appropriate DNA fragment which codes for amino acids 42-649 of

H-SemaL into the vector pMelBacA (4.8 kb Invitrogen) (compare annotation for pMelBacA-H-SEMAL). The cloning took place via BamHI and EcoRI in frame behind the signal sequence present in the vector ("honeybee melittin signal sequence"). A corresponding H-SemaL DNA fragment was amplified using the primer pair h-sema-1 baculo 5' and h-sema-1 baculo 3'.

Primers for amplification (TaKaRa Ex Ta9 polymerase) and cloning:
"h-sema-1 baculo 5'" for amplification without signal sequence and for introducing a BamHI cleavage site
5'-CCGGATCCGCCCAGGGCCACCTAAGGAGCGG-3' (SEQ ID NO: 43)
"h-sema-1 baculo 3'" for amplification without transmembrane domain and for introducing an EcoRI cleavage site
5'-CTGAATTCAGGAGCCAGGGCACAGGCATG-3' (SEQ ID NO: 44).

DETAILED DESCRIPTION OF THE DRAWINGS

Figure 1:
Tissue-specific expression of H-Sema - L

A) Multiple tissue Northern blot (Clontech, Heidelberg, Germany). Loadings from left to right: 2 µg in each lane of Poly-A-RNA from spleen, thymus, prostate, testes, ovaries, small intestine, large intestinal mucosa, peripheral (blood) leukocytes. Size standards are marked.

The blots were hybridized under stringent conditions with an H-SemaL probe 800 base-pairs long.

Figure 2:
Diagrammatic representation of the cloning of the H-SemaL cDNA and of the genomic organization of the H-SemaL encoding sequences (H-SemaL gene)
Top: Location of the EST sequences (accession numbers; location of the EST sequences is shown relative to the AHV-Sema sequence).

Below: Amplified PCR and RACE products and the position of the cDNA clones in relation to the location in the complete H-SemaL cDNA and the open reading frame (ORF) for the encoded protein.
Bottom: Relative position of the exons in the H-SemaL gene in relation to the genomic sequence. The position of the oligonucleotide primer used is indicated by arrows.

Figure 3:
Phylogenetic tree: Obtained by multiple alignment of the listed semaphorin sequences. The phylogenetic relationship of the semaphorins can be deduced from their grouping in the phylogenetic tree.

Figure 4:
FACS analysis of H-SemaL expression in various cell lines and various cell types (compare Example 8).

Figure 5:
Comparative analysis of CD100 and H-SemaL expression (compare Example 9).

Figure 6:
Expression of secretable human SEMA-L (H-SemaL) in HiFive and Sf3 cells (compare Example 10).

Aa 42-649 in pMelBac-A (Invitrogen) in the baculovirus system (Bac-N-Blue, Invitrogen)

Detection with specific chicken antiserum (1:100) and anti-IgY-HRP conjugate (1:3000, from rabbits, Jackson Lab.)
1,4,6 uninfected HiFive cells (serum-free)
2,3,5,7,8 HiFive cells infected with recombinant baculovirus (serum-free)
M Rainbow molecular weight marker (Amersham RPN756)
9,10 infected Sf9 cells (serum-containing medium).

Figure 7: Specificity of the antiserum
Lanes 1-3: chicken 1; lanes 4-6: chicken 2
Lanes 1 and 4: Preimmune serum
Lanes 2 and 5: 60th day of immunization
Lanes 4 and 6: 105th day of immunization

Immunization was carried out with amino acids 179-378 of H-SemaL (with amino-terminal His tag) (compare Example 8, Section 1.)

Figure 8: Depiction of the plasmid map of pMelBacA-H-SEMAL.

The recombinant plasmid was prepared as described in Example 11.

TABLES

Table 1: Various subtypes of semaphorins from various species

Name	Synonym	Species		Reference
H-Sema III	(H-SemaD)	Human	Sec.	(Kolodkin et al. 1993)
CD-100		Human	TM, IC; CD45 associated, expressed in T cells	(Hall et al. 1996)
H-Sema V	(H-SemaA)	Human	Sec.; Locus 3p21.3	(Sekido et al. 1996; Roche et al. 1996)
H-Sema IV	(H-Sema3F)	Human	Sec.; Locus 3p21.3	(Xiang et al. 1996; Sekido et al. 1996)
H-SemaE		Human	Sec.; divergent from M-Sema-E at the 3' end (alignment of reading frame improved)	AB000220 (Yamada 1997 unpublished)
H-SemaK	KIAA0331	Human	Sec.;	(Nagase et al. 1997)
H-SemaL	SEMAL	Human	TM, no IC	This application
M-SemaA		Mouse	Sec.	(Püschel et al. 1995)
M-SemaB		Mouse	TM, IC	(Püschel et al. 1995)
M-SemaC		Mouse	TM, IC	(Püschel et al. 1995)
M-SemaD	M-Sema III	Mouse	Sec.	(Messersmith et al. 1995; Püschel et al. 1995)
M-SemaE		Mouse	Sec.; 5' partial sequence	(Püschel et al. 1995)

Name	Synonym	Species		Reference
M-SemaF1	M-SemaF	Mouse	TM, IC	(Inagaki et al. 1995)
M-SemaG2	M-SemaG	Mouse	TM, IC; expressed in lymphoid cells, mouse homolog of CD100	(Furuyama et al. 1996)
M-SemaF2	M-SemaF	Mouse	TM, IC; Thrombospondin motif	(Adams et al. 1996)
M-SemaG1	M-SemaG	Mouse	TM, IC; Thrombospondin motif	(Adams et al. 1996)
M-SemaH		Mouse	Sec.	(Christensen 1996 unpub) Z80941
M-Sema VIa		Mouse	TM, IC	(Zhou et al. 1997)
M-SemaL	SemaI	Mouse	Partial sequence	This application
Collapsin-1		Chicken	Sec.	(Luo et al. 1993)
Collapsin-2		Chicken	Sec.	(Luo et al. 1995)
Collapsin-3		Chicken	Sec.	(Luo et al. 1995)
Collapsin-4		Chicken	Partial sequence	(Luo et al. 1995)
Collapsin-5		Chicken	Sec.	(Luo et al. 1995)
R-Sema III		Rat	Sec.	(Giger et al. 1996)

Name	Synonym	Species		Reference
T-Sema I		Tribolium confusum	TM, IC	(Kolodkin et al. 1993)
Ce-Semal		C.elegans	TM, IC	U15667 (Roy1994 unpublished)
G-Sema I	Fasciclin-IV	Grasshopper	TM, IC	(Kolodkin et al. 1992)
D-Sema I		Drosophila	TM, IC	(Kolodkin et al. 1993)
D-Sema II		Drosophila	Sec.	(Kolodkin et al. 1993)
AHV-Sema		AHV-1	Sec.	(Ensser and Fleckenstein, 1995)
ORF-A39		Vaccinia	Sec.	(Kolodkin et al. 1993)
ORF-A39 homologous		Variola	Sec.;	(Kolodkin et al. 1993)

TM: transmembrane domain

Sec.: secreted

IC: presumably intracellular cytoplasmic sequence motif

Table 2: cDNA sequence of H-SemaL (2636 nucleotides) (SEQ ID NO.: 1)

```
1     cggggccacg ggatgacgcc tcctccgccc ggacgtgccg cccccagcgc
51    accgcgcgcc cgcgtccctg gcccgccggc tcggttgggg cttccgctgc
101   ggctgcggct gctgctgctg ctctgggcgg ccgccgcctc cgcccagggc
151   cacctaagga gcggaccccg catcttcgcc gtctggaaag gccatgtagg
201   gcaggaccgg gtggactttg gccagactga gccgcacacg gtgcttttcc
251   acgagccagg cagctcctct gtgtgggtgg gaggacgtgg caaggtctac
301   ctctttgact tccccgaggg caagaacgca tctgtgcgca cggtgaatat
351   cggctccaca aaggggtcct gtctggataa gcgggactgc gagaactaca
401   tcactctcct ggagaggcgg agtgaggggc tgctggcctg tggcaccaac
451   gcccggcacc ccagctgctg gaacctggtg aatggcactg tggtgccact
501   tggcgagatg agaggctacg ccccttcag cccggacgag aactccctgg
551   ttctgtttga aggggacgag gtgtattcca ccatccggaa gcaggaatac
601   aatgggaaga tccctcggtt ccgccgcatc cggggcgaga gtgagctgta
651   caccagtgat actgtcatgc agaacccaca gttcatcaaa gccaccatcg
701   tgcaccaaga ccaggcttac gatgacaaga tctactactt cttccgagag
751   gacaatcctg acaagaatcc tgaggctcct ctcaatgtgt cccgtgtggc
801   ccagttgtgc agggggggacc agggtgggga aagttcactg tcagtctcca
851   agtggaacac ttttctgaaa gccatgctgg tatgcagtga tgctgccacc
901   aacaagaact tcaacaggct gcaagacgtc ttcctgctcc ctgaccccag
951   cggccagtgg agggacacca gggtctatgg tgttttctcc aacccctgga
1001  actactcagc cgtctgtgtg tattccctcg gtgacattga caaggtcttc
1051  cgtacctcct cactcaaggg ctaccactca agccttccca acccgcggcc
1101  tggcaagtgc ctcccagacc agcagccgat acccacagag accttccagg
1151  tggctgaccg tcacccagag gtggcgcaga gggtggagcc catggggcct
1201  ctgaagacgc cattgttcca ctctaaatac cactaccaga aagtggccgt
1251  tcaccgcatg caagccagcc acggggagac ctttcatgtg ctttacctaa
1301  ctacagacag gggcactatc cacaaggtgg tggaaccggg ggagcaggag
1351  cacagcttcg ccttcaacat catggagatc cagcccttcc gccgcgcggc
1401  tgccatccag accatgtcgc tggatgctga gcggaggaag ctgtatgtga
1451  gctcccagtg ggaggtgagc caggtgcccc tggacctgtg tgaggtctat
1501  ggcgggggct gccacggttg cctcatgtcc cgagacccct actgcggctg
1551  ggaccagggc cgctgcatct ccatctacag ctccgaacgg tcagtgctgc
1601  aatccattaa tccagccgag ccacacaagg agtgtcccaa ccccaaacca
```

1651 gacaaggccc cactgcagaa ggtttccctg gccccaaact ctcgctacta
1701 cctgagctgc cccatggaat cccgccacgc cacctactca tggcgccaca
1751 aggagaacgt ggagcagagc tgcgaacctg gtcaccagag ccccaactgc
1801 atcctgttca tcgagaacct cacggcgcag cagtacggcc actacttctg
1851 cgaggcccag gagggctcct acttccgcga ggctcagcac tggcagctgc
1901 tgcccgagga cggcatcatg gccgagcacc tgctgggtca tgcctgtgcc
1951 ctggctgcct ccctctggct gggggtgctg cccacactca ctcttggctt
2001 gctggtccac tagggcctcc cgaggctggg catgcctcag gcttctgcag
2051 cccagggcac tagaacgtct cacactcaga gccggctggc ccgggagctc
2101 cttgcctgcc acttcttcca ggggacagaa taacccagtg gaggatgcca
2151 ggcctggaga cgtccagccg caggcggctg ctgggcccca ggtggcgcac
2201 ggatggtgag ggctgagaa tgagggcacc gactgtgaag ctggggcatc
2251 gatgacccaa gactttatct tctggaaaat atttttcaga ctcctcaaac
2301 ttgactaaat gcagcgatgc tcccagccca agagcccatg ggtcggggag
2351 tgggtttgga taggagagct gggactccat ctcgaccctg gggctgaggc
2401 ctgagtcctt ctggactctt ggtacccaca ttgcctcctt cccctccctc
2451 tctcatggct gggtggctgg tgttcctgaa gacccagggc taccctctgt
2501 ccagccctgt cctctgcagc tccctctctg gtcctgggtc ccacaggaca
2551 gccgccttgc atgtttattg aaggatgttt gctttccgga cggaaggacg
2601 gaaaaagctc tgaaaaaaaa aaaaaaaaaa aaaaaa

Table 3: Nucleotide sequence of the cDNA of M-SemaL (partial, 1195 nucleotides) (SEQ ID NO.: 2)

1 cggggctgcg ggatgacgcc tcctcctccc ggacgtgccg cccccagcgc
51 accgcgcgcc cgcgtcctca gcctgccggc tcggttcggg ctcccgctgc
101 ggctgcggct tctgctggtg ttctgggtgg ccgccgcctc cgcccaaggc
151 cactcgagga gcggaccccg catctccgcc gtctggaaag ggcaggacca
201 tgtggacttt agccagcctg agccacacac cgtgcttttc catgagccgg
251 gcagcttctc tgtctgggtg ggtggacgtg gcaaggtcta ccacttcaac
301 ttccccgagg gcaagaatgc ctctgtgcgc acggtgaaca tcggctccac
351 aaaggggtcc tgtcaggaca aacaggactg tgggaattac atcactcttc
401 tagaaaggcg ggtaatggg ctgctggtct gtggcaccaa tgcccggaag
451 cccagctgct ggaacttggt gaatgacagt gtggtgatgt cacttggtga

501 gatgaaaggc tatgcccct tcagcccgga tgagaactcc ctggttctgt
551 ttgaaggaga tgaagtgtac tctaccatcc ggaagcagga atacaacggg
601 aagatccctc ggtttcgacg cattcggggc gagagtgaac tgtacacaag
651 tgatacagtc atgcagaacc cacagttcat caaggccacc attgtgcacc
701 aagaccaagc ctatgatgat aagatctact acttcttccg agaagacaac
751 cctgacaaga accccgaggc tcctctcaat gtgtcccgag tagcccagtt
801 gtgcaggggg gaccagggtg gtgagagttc gttgtctgtc tccaagtgga
851 acaccttcct gaaagccatg ttggtctgca gcgatgcagc caccaacagg
901 aacttcaatc ggctgcaaga tgtcttcctg ctccctgacc ccagtggcca
951 gtggagagat accagggtct atggcgtttt ctccaacccc tggaactact
1001 cagctgtctg cgtgtattcg cttggtgaca ttgacagagt cttccgtacc
1051 tcatcgctca aaggctacca catgggcctt tccaaccctc gacctggcat
1101 gtgcctccca aaaaagcagc ccatacccac agaaaccttc caggtagctg
1151 atagtcaccc agaggtggct cagagggtgg aacctatggg gcccc

Table 4: Amino acid sequence of H-SemaL (666 amino acids) (SEQ ID NO.: 3)

1 MTPPPPGRAA PSAPRARVPG PPARLGLPLR LRLLLLLWAA AASAQGHLRS
51 GPRIFAVWKG HVGQDRVDFG QTEPHTVLFH EPGSSSVWVG GRGKVYLFDF
101 PEGKNASVRT VNIGSTKGSC LDKRDCENYI TLLERRSEGL LACGTNARHP
151 SCWNLVNGTV VPLGEMRGYA PFSPDENSLV LFEGDEVYST IRKQEYNGKI
201 PRFRRIRGES ELYTSDTVMQ NPQFIKATIV HQDQAYDDKI YYFFREDNPD
251 KNPEAPLNVS RVAQLCRGDQ GGESSLSVSK WNTFLKAMLV CSDAATNKNF
301 NRLQDVFLLP DPSGQWRDTR VYGVFSNPWN YSAVCVYSLG DIDKVFRTSS
351 LKGYHSSLPN PRPGKCLPDQ QPIPTETFQV ADRHPEVAQR VEPMGPLKTP
401 LFHSKYHYQK VAVHRMQASH GETFHVLYLT TDRGTIHKVV EPGEQEHSFA
451 FNIMEIQPFR RAAAIQTMSL DAERRKLYVS SQWEVSQVPL DLCEVYGGGC
501 HGCLMSRDPY CGWDQGRCIS IYSSERSVLQ SINPAEPHKE CPNPKPDKAP
551 LQKVSLAPNS RYYLSCPMES RHATYSWRHK ENVEQSCEPG HQSPNCILFI
601 ENLTAQQYGH YFCEAQEGSY FREAQHWQLL PEDGIMAEHL LGHACALAAS
651 LWLGVLPTLT LGLLVH

Table 5: (Partial) amino acid sequence of M-SemaL (394 amino acids, corresponding to position 1-396 of H-SemaL) (SEQ ID NO.: 4)

1	MTPPPPGRAA	PSAPRARVLS LPARFGLPLR LRLLLVFWVA AASAQGHSRS
51	GPRISAVWKG	QDHVDFSQPE PHTVLFHEPG SFSVWVGGRG KVYHFNFPEG
101	KNASVRTVNI	GSTKGSCQDK QDCGNYITLL ERRGNGLLVC GTNARKPSCW
151	NLVNDSVVMS	LGEMKGYAPF SPDENSLVLF EGDEVYSTIR KQEYNGKIPR
201	FRRIRGESEL	YTSDTVMQNP QFIKATIVHQ DQAYDDKIYY FFREDNPDKN
251	PEAPLNVSRV	AQLCRGDQGG ESSLSVSKWN TFLKAMLVCS DAATNRNFNR
301	LQDVFLLPDP	SGQWRDTRVY GVFSNPWNYS AVCVYSLGDI DRVFRTSSLK
351	GYHMGLSNPR	PGMCLPKKQP IPTETFQVAD SHPEVAQRVE PMGP

Table 6: Synthetic oligonucleotides (Eurogentec, Seraing, Belgium)

Number of the primer/name	Nucleotide sequence of the primer (of the synthetic oligonucleotides)	
91506/AP2	actcactatagggctcgagcggc	(SEQ ID NO.: 5)
121234	agccgcacacggtgcttttc	(SEQ ID NO.: 6)
121235/Est 2	gcacagatgcgttcttgccc	(SEQ ID NO.: 7)
121236/Est 3	accatagaccctggtgtccc	(SEQ ID NO.: 8)
121237/Est 4	gcagtgatgctgccaccaac	(SEQ ID NO.: 9)
121238	ccagaccatgtcgctggatg	(SEQ ID NO.: 10)
121239/Est 6	acatgaggcaaccgtggcag	(SEQ ID NO.: 11)
131989/AP1	ccatcctaatacgactcactatagggc	(SEQ ID NO.: 12)
131990/Est 7	aggtagaccttgccacgtcc	(SEQ ID NO.: 13)
131991	gaacttcaacaggctgcaagacg	(SEQ ID NO.: 14)
131992	atgctgagcggaggaagctg	(SEQ ID NO.: 15)
131993	ccgccatacacctcacacag	(SEQ ID NO.: 16)
150788	ctggaagctttctgtgggtatcggctgc	(SEQ ID NO.: 17)
150789	tttggatccctggttctgtttgaag	(SEQ ID NO.: 18)
167579/cDNA	ttctagaattcagcggccgctttttttttttttttttttttttttttttttvn	(SEQ ID NO.: 19)
Synthesis primer		
168421	ggggaaagttcactgtcagtctccaag	(SEQ ID NO.: 20)
168422	gggaatacacacagacggctgagtag	(SEQ ID NO.: 21)

207608/	agcaagttcagcctggttaagt	(SEQ ID NO.: 22)
Amplification of λgt10 insert		
207609/	ttatgagtatttcttccaggg	(SEQ ID NO.: 23)
Amplification of λgt10 insert		
232643/Est 13	ccattaatccagccgagccacacaag	(SEQ ID NO.: 24)
232644/Est 14	catctacagctccgaacggtcagtg	(SEQ ID NO.: 25)
233084	cagcggaagccccaaccgag	(SEQ ID NO.: 26)
240655/hs 5	gggatgacgcctcctccgcccgg	(SEQ ID NO.: 27)
240656/hs 3	aagcttcacgtggaccagcaagccaagagtg	(SEQ ID NO.: 28)
240657/hs 3c	aagctttttccgtccttccgtccgg	(SEQ ID NO.: 29)
243068	atggtgagcaagggcgaggagctg	(SEQ ID NO.: 30)
243069	cttgtacagctcgtccatgccgag	(SEQ ID NO.: 31)
260812	GGGTGGTGAGAGTTCGTTGTCTGTC	(SEQ ID NO.: 32)
260813	GAGCGATGAGGTACGGAAGACTCTG	(SEQ ID NO.: 33)

Table 7: Nucleotide sequence of the recombinant plasmid pCR2.1-H-SemaL (SEQ ID NO.: 34)

```
  1 AGCGCCCAAT ACGCAAACCG CCTCTCCCCG CGCGTTGGCC GATTCATTAA
 51 TGCAGCTGGC ACGACAGGTT TCCCGACTGG AAAGCGGGCA GTGAGCGCAA
101 CGCAATTAAT GTGAGTTAGC TCACTCATTA GGCACCCCAG GCTTTACACT
151 TTATGCTTCC GGCTCGTATG TTGTGTGGAA TTGTGAGCGG ATAACAATTT
201 CACACAGGAA ACAGCTATGA CCATGATTAC GCCaagcttc acgtggacca
251 gcaagccaag agtgagtgtg ggcagcaccc ccagccagag ggaggcagcc
301 agggcacagg catgacccag caggtgctcg gccatgatgc cgtcctcggg
351 cagcagctgc cagtgctgag cctcgcggaa gtaggagccc tcctgggcct
401 cgcagaagta gtggccgtac tgctgcgccg tgaggttctc gatgaacagg
451 atgcagttgg ggctctggtg accaggttcg cagctctgct ccacgttctc
501 cttgtggcgc catgagtagg tggcgtggcg ggattccatg gggcagctca
551 ggtagtagcg agagtttggg gccagggaaa ccttctgcag tggggccttg
601 tctggtttgg ggttgggaca ctccttgtgt ggctcggctg gattaatgga
651 ttgcagcact gaccgttcgg agctgtagat ggagatgcag cggccctggt
701 cccagccgca gtaggggtct cgggacatga ggcaaccgtg gcagcccccg
751 ccatagacct cacacaggtc caggggcacc tggctcacct cccactggga
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801 gctcacatac agcttcctcc gctcagcatc cagcgacatg gtctggatgg
851 cagccgcgcg gcggaagggc tggatctcca tgatgttgaa ggcgaagctg
901 tgctcctgct ccccccggttc caccaccttg tggatagtgc ccctgtctgt
951 agttaggtaa agcacatgaa aggtctcccc gtggctggct tgcatgcggt
1001 gaacggccac tttctggtag tggtatttag agtggaacaa tggcgtcttc
1051 agaggcccca tgggctccac cctctgcgcc acctctgggt gacggtcagc
1101 cacctggaag gtctctgtgg gtatcggctg ctggtctggg aggcacttgc
1151 caggccgcgg gttgggaagg cttgagtggt agcccttgag tgaggaggta
1201 cggaagacct tgtcaatgtc accgagggaa tacacacaga cggctgagta
1251 gttccagggg ttggagaaaa caccatagac cctggtgtcc ctccactggc
1301 cgctggggtc agggagcagg aagacgtctt gcagcctgtt gaagttcttg
1351 ttggtggcag catcactgca taccagcatg gctttcagaa aagtgttcca
1401 cttggagact gacagtgaac tttccccacc ctggtccccc ctgcacaact
1451 gggccacacg ggacacattg agaggagcct caggattctt gtcaggattg
1501 tcctctcgga agaagtagta gatcttgtca tcgtaagcct ggtcttggtg
1551 cacgatggtg gctttgatga actgtgggtt ctgcatgaca gtatcactgg
1601 tgtacagctc actctcgccc cggatgcggc ggaaccgagg gatcttccca
1651 ttgtattcct gcttccggat ggtggaatac acctcgtccc cttcaaacag
1701 aaccagggag ttctcgtccg ggctgaaggg ggcgtagcct ctcatctcgc
1751 caagtggcac cacagtgcca ttcaccaggt tccagcagct ggggtgccgg
1801 gcgttggtgc cacaggccag cagcccctca ctccgcctct ccaggagagt
1851 gatgtagttc tcgcagtccc gcttatccag acaggacccc tttgtggagc
1901 cgatattcac cgtgcgcaca gatgcgttct tgccctcggg gaagtcaaag
1951 aggtagacct tgccacgtcc tcccacccac acagaggagc tgcctggctc
2001 gtggaaaagc accgtgtgcg gctcagtctg gccaaagtcc acccggtcct
2051 gccctacatg gcctttccag acggcgaaga tgcggggtcc gctccttagg
2101 tggccctggg cggaggcggc ggccgcccag agcagcagca gcagccgcag
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2201 gtgcgctggg ggcggcacgt ccgggcggag gaggcgtcat cccaagccga
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2301 GGCCCAATTC GCCCTATAGT GAGTCGTATT ACAATTCACT GGCCGTCGTT
2351 TTACAACGTC GTGACTGGGA AAACCCTGGC GTTACCCAAC TTAATCGCCT
2401 TGCAGCACAT CCCCCTTTCG CCAGCTGGCG TAATAGCGAA GAGGCCCGCA
2451 CCGATCGCCC TTCCCAACAG TTGCGCAGCC TGAATGGCGA ATGGGACGCG
2501 CCCTGTAGCG GCGCATTAAG CGCGGCGGGT GTGGTGGTTA CGCGCAGCGT

2551 GACCGCTACA CTTGCCAGCG CCCTAGCGCC CGCTCCTTTC GCTTTCTTCC
2601 CTTCCTTTCT CGCCACGTTC GCCGGCTTTC CCCGTCAAGC TCTAAATCGG
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3001 GCTATCTGGA CAAGGGAAAA CGCAAGCGCA AAGAGAAAGC AGGTAGCTTG
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3651 AGCGAAACAT CGCATCGAGC GAGCACGTAC TCGGATGGAA GCCGGTCTTG
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3751 CTGTTCGCCA GGCTCAAGGC GCGCATGCCC GACGGCGAGG ATCTCGTCGT
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4401 ACAGTAAGAG AATTATGCAG TGCTGCCATA ACCATGAGTG ATAACACTGC
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4751 ATCTGGAGCC GGTGAGCGTG GGTCTCGCGG TATCATTGCA GCACTGGGGC
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5401 GCTGAACGGG GGGTTCGTGC ACACAGCCCA GCTTGGAGCG AACGACCTAC
5451 ACCGAACTGA GATACCTACA GCGTGAGCAT TGAGAAAGCG CCACGCTTCC
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5551 GAGAGCGCAC GAGGGAGCTT CCAGGGGGAA ACGCCTGGTA TCTTTATAGT
5601 CCTGTCGGGT TTCGCCACCT CTGACTTGAG CGTCGATTTT TGTGATGCTC
5651 GTCAGGGGGG CGGAGCCTAT GGAAAAACGC CAGCAACGCG GCCTTTTTAC
5701 GGTTCCTGGC CTTTTGCTGG CCTTTTGCTC ACATGTTCTT TCCTGCGTTA
5751 TCCCCTGATT CTGTGGATAA CCGTATTACC GCCTTTGAGT GAGCTGATAC
5801 CGCTCGCCGC AGCCGAACGA CCGAGCGCAG CGAGTCAGTG
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5851 CGGAAG

Table 8: Nucleotide sequence of the recombinant expression plasmid pCDNA3.1(-)H-SemaL-MycHisA (SEQ ID NO.: 35)

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 101 GGAGGTCGCT GAGTAGTGCG CGAGCAAAAT TTAAGCTACA ACAAGGCAAG
 151 GCTTGACCGA CAATTGCATG AAGAATCTGC TTAGGGTTAG GCGTTTTGCG
 201 CTGCTTCGCG ATGTACGGGC CAGATATACG CGTTGACATT GATTATTGAC
 251 TAGTTATTAA TAGTAATCAA TTACGGGGTC ATTAGTTCAT AGCCCATATA
 301 TGGAGTTCCG CGTTACATAA CTTACGGTAA ATGGCCCGCC TGGCTGACCG
 351 CCCAACGACC CCCGCCCATT GACGTCAATA ATGACGTATG TTCCCATAGT
 401 AACGCCAATA GGGACTTTCC ATTGACGTCA ATGGGTGGAC TATTTACGGT
 451 AAACTGCCCA CTTGGCAGTA CATCAAGTGT ATCATATGCC AAGTACGCCC
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 601 TCGCTATTAC CATGGTGATG CGGTTTTGGC AGTACATCAA TGGGCGTGGA
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 701 TGGGAGTTTG TTTTGGCACC AAAATCAACG GGACTTTCCA AAATGTCGTA
 751 ACAACTCCGC CCCATTGACG CAAATGGGCG GTAGGCGTGT
     ACGGTGGGAG
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3801 GGAAAGTCCC CAGGCTCCCC AGGCAGGCAG AAGTATGCAA AGCATGCATC
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5051 CTTCTATGAA AGGTTGGGCT TCGGAATCGT TTTCCGGGAC GCCGGCTGGA
5101 TGATCCTCCA GCGCGGGGAT CTCATGCTGG AGTTCTTCGC CCACCCCAAC
5151 TTGTTTATTG CAGCTTATAA TGGTTACAAA TAAAGCAATA GCATCACAAA
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7301 AAAGGGAATA AGGGCGACAC GGAAATGTTG AATACTCATA CTCTTCCTTT
7351 TTCAATATTA TTGAAGCATT TATCAGGGTT ATTGTCTCAT GAGCGGATAC
7401 ATATTTGAAT GTATTTAGAA AAATAAACAA ATAGGGGTTC CGCGCACATT
7451 TCCCCGAAAA GTGCCACCTG ACGTC

Table 9: Nucleotide sequence of the recombinant plasmid pcDNA3.1-H-SemaL-EGFP-MychisA (SEQ ID NO.: 36)

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51 TGCTCTGATG CCGCATAGTT AAGCCAGTAT CTGCTCCCTG CTTGTGTGTT
101 GGAGGTCGCT GAGTAGTGCG CGAGCAAAAT TTAAGCTACA ACAAGGCAAG
151 GCTTGACCGA CAATTGCATG AAGAATCTGC TTAGGGTTAG GCGTTTTGCG
201 CTGCTTCGCG ATGTACGGGC CAGATATACG CGTTGACATT GATTATTGAC
251 TAGTTATTAA TAGTAATCAA TTACGGGGTC ATTAGTTCAT AGCCCATATA
301 TGGAGTTCCG CGTTACATAA CTTACGGTAA ATGGCCCGCC TGGCTGACCG
351 CCCAACGACC CCCGCCCATT GACGTCAATA ATGACGTATG TTCCCATAGT
401 AACGCCAATA GGGACTTTCC ATTGACGTCA ATGGGTGGAC TATTTACGGT
451 AAACTGCCCA CTTGGCAGTA CATCAAGTGT ATCATATGCC AAGTACGCCC
501 CCTATTGACG TCAATGACGG TAAATGGCCC GCCTGGCATT ATGCCCAGTA
551 CATGACCTTA TGGGACTTTC CTACTTGGCA GTACATCTAC GTATTAGTCA
601 TCGCTATTAC CATGGTGATG CGGTTTTGGC AGTACATCAA TGGGCGTGGA
651 TAGCGGTTTG ACTCACGGGG ATTTCCAAGT CTCCACCCCA TTGACGTCAA
701 TGGGAGTTTG TTTTGGCACC AAAATCAACG GGACTTTCCA AAATGTCGTA
751 ACAACTCCGC CCCATTGACG CAAATGGGCG GTAGGCGTGT ACGGTGGGAG
801 GTCTATATAA GCAGAGCTCT CTGGCTAACT AGAGAACCCA CTGCTTACTG

851 GCTTATCGAA ATTAATACGA CTCACTATAG GGAGACCCAA GCTGGCTAGC
901 GTTTAAACGG GCCCTCTAGA CTCGAGCGGC CGCCACTGTG CTGGATATCT
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GCTGACCCTG
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3251 GAGCGCACCA TCTTCTTCAA GGACGACGGC AACTACAAGA CCCGCGCCGA
3301 GGTGAAGTTC GAGGGCGACA CCCTGGTGAA CCGCATCGAG CTGAAGGGCA
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3651 GGATCACTCT CGGCATGGAC GAGCTGTACA Aggtgaagct tGGGCCCGAA
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4251 CCTCGACCCC AAAAAACTTG ATTAGGGTGA TGGTTCACGT AGTGGGCCAT

4301 CGCCCTGATA GACGGTTTTT CGCCCTTTGA CGTTGGAGTC CACGTTCTTT
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5801 CCGGGACGCC GGCTGGATGA TCCTCCAGCG CGGGGATCTC ATGCTGGAGT
5851 TCTTCGCCCA CCCCAACTTG TTTATTGCAG CTTATAATGG TTACAAATAA
5901 AGCAATAGCA TCACAAATTT CACAAATAAA GCATTTTTTT CACTGCATTC
5951 TAGTTGTGGT TTGTCCAAAC TCATCAATGT ATCTTATCAT GTCTGTATAC
6001 CGTCGACCTC TAGCTAGAGC TTGGCGTAAT CATGGTCATA GCTGTTTCCT

6051 GTGTGAAATT GTTATCCGCT CACAATTCCA CACAACATAC GAGCCGGAAG
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6151 TTGCGTTGCG CTCACTGCCC GCTTTCCAGT CGGGAAACCT GTCGTGCCAG
6201 CTGCATTAAT GAATCGGCCA ACGCGCGGGG AGAGGCGGTT TGCGTATTGG
6251 GCGCTCTTCC GCTTCCTCGC TCACTGACTC GCTGCGCTCG GTCGTTCGGC
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6601 TTCTCCCTTC GGGAAGCGTG GCGCTTTCTC AATGCTCACG CTGTAGGTAT
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6701 CCCCGTTCAG CCCGACCGCT GCGCCTTATC CGGTAACTAT CGTCTTGAGT
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6801 AGGATTAGCA GAGCGAGGTA TGTAGGCGGT GCTACAGAGT TCTTGAAGTG
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6901 TGCTGAAGCC AGTTACCTTC GGAAAAAGAG TTGGTAGCTC TTGATCCGGC
6951 AAACAAACCA CCGCTGGTAG CGGTGGTTTT TTTGTTTGCA AGCAGCAGAT
7001 TACGCGCAGA AAAAAAGGAT CTCAAGAAGA TCCTTTGATC TTTTCTACGG
7051 GGTCTGACGC TCAGTGGAAC GAAAACTCAC GTTAAGGGAT TTTGGTCATG
7101 AGATTATCAA AAAGGATCTT CACCTAGATC CTTTTAAATT AAAAATGAAG
7151 TTTTAAATCA ATCTAAAGTA TATATGAGTA AACTTGGTCT GACAGTTACC
7201 AATGCTTAAT CAGTGAGGCA CCTATCTCAG CGATCTGTCT ATTTCGTTCA
7251 TCCATAGTTG CCTGACTCCC CGTCGTGTAG ATAACTACGA TACGGGAGGG
7301 CTTACCATCT GGCCCCAGTG CTGCAATGAT ACCGCGAGAC CCACGCTCAC
7351 CGGCTCCAGA TTTATCAGCA ATAAACCAGC CAGCCGGAAG GGCCGAGCGC
7401 AGAAGTGGTC CTGCAACTTT ATCCGCCTCC ATCCAGTCTA TTAATTGTTG
7451 CCGGGAAGCT AGAGTAAGTA GTTCGCCAGT TAATAGTTTG CGCAACGTTG
7501 TTGCCATTGC TACAGGCATC GTGGTGTCAC GCTCGTCGTT TGGTATGGCT
7551 TCATTCAGCT CCGGTTCCCA ACGATCAAGG CGAGTTACAT GATCCCCCAT
7601 GTTGTGCAAA AAAGCGGTTA GCTCCTTCGG TCCTCCGATC GTTGTCAGAA
7651 GTAAGTTGGC CGCAGTGTTA TCACTCATGG TTATGGCAGC ACTGCATAAT
7701 TCTCTTACTG TCATGCCATC CGTAAGATGC TTTTCTGTGA CTGGTGAGTA
7751 CTCAACCAAG TCATTCTGAG AATAGTGTAT GCGGCGACCG AGTTGCTCTT

7801 GCCCGGCGTC AATACGGGAT AATACCGCGC CACATAGCAG AACTTTAAAA
7851 GTGCTCATCA TTGGAAAACG TTCTTCGGGG CGAAAACTCT CAAGGATCTT
7901 ACCGCTGTTG AGATCCAGTT CGATGTAACC CACTCGTGCA CCCAACTGAT
7951 CTTCAGCATC TTTTACTTTC ACCAGCGTTT CTGGGTGAGC AAAAACAGGA
8001 AGGCAAAATG CCGCAAAAAA GGGAATAAGG GCGACACGGA AATGTTGAAT
8051 ACTCATACTC TTCCTTTTTC AATATTATTG AAGCATTTAT CAGGGTTATT
8101 GTCTCATGAG CGGATACATA TTTGAATGTA TTTAGAAAAA TAAACAAATA
8151 GGGGTTCCGC GCACATTTCC CCGAAAAGTG CCACCTGACG TC

Table10: Nucleotide sequence of the recombinant plasmid pIND-H-SemaL-EE (SEQ ID NO.:37)

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  51 TGTTCTCGTT AGCTCGATGG ACAAGTGCAT TGTTCTCTTG CTGAAAGCTC
 101 GATGGACAAG TGCATTGTTC TCTTGCTGAA AGCTCGATGG ACAAGTGCAT
 151 TGTTCTCTTG CTGAAAGCTC AGTACCCGGG AGTACCCTCG ACCGCCGGAG
 201 TATAAATAGA GGCGCTTCGT CTACGGAGCG ACAATTCAAT TCAAACAAGC
 251 AAAGTGAACA CGTCGCTAAG CGAAAGCTAA GCAAATAAAC AAGCGCAGCT
 301 GAACAAGCTA AACAATCTGC AGTAAAGTGC AAGTTAAAGT GAATCAATTA
 351 AAAGTAACCA GCAACCAAGT AAATCAACTG CAACTACTGA AATCTGCCAA
 401 GAAGTAATTA TTGAATACAA GAAGAGAACT CTGAATACTT TCAACAAGTT
 451 ACCGAGAAAG AAGAACTCAC ACACAGCTAG CGTTTAAACT TAAGCTTGGT
 501 ACCGAGCTCG GATCCACTAG TCCAGTGTGG TGgaattcgg cttgggatga
 551 cgcctcctcc gcccggacgt gccgccccca gcgcaccgcg cgcccgcgtc
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2801 GCTCTATGGC TTCTGAGGCG GAAAGAACCA GCTGGGGCTC TAGGGGGTAT
2851 CCCCACGCGC CCTGTAGCGG CGCATTAAGC GCGGCGGGTG TGGTGGTTAC
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3251 ATGAGCTGAT TTAACAAAAA TTTAACGCGA ATTAATTCTG TGGAATGTGT
3301 GTCAGTTAGG GTGTGGAAAG TCCCCAGGCT CCCCAGGCAG GCAGAAGTAT

3351 GCAAAGCATG CATCTCAATT AGTCAGCAAC CAGGTGTGGA AAGTCCCCAG
3401 GCTCCCCAGC AGGCAGAAGT ATGCAAAGCA TGCATCTCAA TTAGTCAGCA
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4051 ATGCGGCGGC TGCATACGCT TGATCCGGCT ACCTGCCCAT TCGACCACCA
4101 AGCGAAACAT CGCATCGAGC GAGCACGTAC TCGGATGGAA GCCGGTCTTG
4151 TCGATCAGGA TGATCTGGAC GAAGAGCATC AGGGGCTCGC GCCAGCCGAA
4201 CTGTTCGCCA GGCTCAAGGC GCGCATGCCC GACGGCGAGG ATCTCGTCGT
4251 GACCCATGGC GATGCCTGCT TGCCGAATAT CATGGTGGAA AATGGCCGCT
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4751 TTGTGGTTTG TCCAAACTCA TCAATGTATC TTATCATGTC TGTATACCGT
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6801 CAAAATGCCG CAAAAAAGGG AATAAGGGCG ACACGGAAAT GTTGAATACT

6851 CATACTCTTC CTTTTTCAAT ATTATTGAAG CATTTATCAG GGTTATTGTC
6901 TCATGAGCGG ATACATATTT GAATGTATTT AGAAAAATAA ACAAATAGGG
6951 GTTCCGCGCA CATTTCCCCG AAAAGTGCCA CCTGACGTCG ACGGATCGGG

Table11: Nucleotide sequence of the recombinant plasmid pIND-H-SemaL-EA (SEQ ID NO.:38)

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51 TGTTCTCGTT AGCTCGATGG ACAAGTGCAT TGTTCTCTTG CTGAAAGCTC
101 GATGGACAAG TGCATTGTTC TCTTGCTGAA AGCTCGATGG ACAAGTGCAT
151 TGTTCTCTTG CTGAAAGCTC AGTACCCGGG AGTACCCTCG ACCGCCGGAG
201 TATAAATAGA GGCGCTTCGT CTACGGAGCG ACAATTCAAT TCAAACAAGC
251 AAAGTGAACA CGTCGCTAAG CGAAAGCTAA GCAAATAAAC AAGCGCAGCT
301 GAACAAGCTA AACAATCTGC AGTAAAGTGC AAGTTAAAGT GAATCAATTA
351 AAAGTAACCA GCAACCAAGT AAATCAACTG CAACTACTGA AATCTGCCAA
401 GAAGTAATTA TTGAATACAA GAAGAGAACT CTGAATACTT TCAACAAGTT
451 ACCGAGAAAG AAGAACTCAC ACACAGCTAG CGTTTAAACT TAAGCTTGGT
501 ACCGAGCTCG GATCCACTAG TCCAGTGTGG TGgaattcgg cttgggatga
551 cgcctcctcc gcccggacgt gccgccccca gcgcaccgcg cgcccgcgtc
601 cctggcccgc cggctcggtt ggggcttccg ctgcggctgc ggctgctgct
651 gctgctctgg gcggccgccg cctccgccca gggccaccta aggagcggac
701 cccgcatctt cgccgtctgg aaaggccatg tagggcagga ccgggtggac
751 tttggccaga ctgagccgca cacggtgctt ttccacgagc caggcagctc
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1001 gctggaacct ggtgaatggc actgtggtgc cacttggcga gatgagaggc
1051 tacgcccccc tcagcccgga cgagaactcc ctggttctgt ttgaagggga
1101 cgaggtgtat tccaccatcc ggaagcagga atacaatggg aagatccctc
1151 ggttccgccg catccggggc gagagtgagc tgtacaccag tgatactgtc
1201 atgcagaacc cacagttcat caaagccacc atcgtgcacc aagaccaggc
1251 ttacgatgac aagatctact acttcttccg agaggacaat cctgacaaga
1301 atcctgaggc tcctctcaat gtgtcccgtg tggcccagtt gtgcaggggg

1351 gaccagggtg gggaaagttc actgtcagtc tccaagtgga acacttttct
1401 gaaagccatg ctggtatgca gtgatgctgc caccaacaag aacttcaaca
1451 ggctgcaaga cgtcttcctg ctccctgacc ccagcggcca gtggagggac
1501 accagggtct atggtgtttt ctccaacccc tggaactact cagccgtctg
1551 tgtgtattcc ctcggtgaca ttgacaaggt cttccgtacc tcctcactca
1601 agggctacca ctcaagcctt cccaacccgc ggcctggcaa gtgcctccca
1651 gaccagcagc cgatacccac agagaccttc caggtggctg accgtcaccc
1701 agaggtggcg cagagggtgg agcccatggg gcctctgaag acgccattgt
1751 tccactctaa ataccactac cagaaagtgg ccgttcaccg catgcaagcc
1801 agccacgggg agacctttca tgtgctttac ctaactacag acaggggcac
1851 tatccacaag gtggtggaac cgggggagca ggagcacagc ttcgccttca
1901 acatcatgga gatccagccc ttccgccgcg cggctgccat ccagaccatg
1951 tcgctggatg ctgagcggag gaagctgtat gtgagctccc agtgggaggt
2001 gagccaggtg cccctggacc tgtgtgaggt ctatggcggg ggctgccacg
2051 gttgcctcat gtcccgagac ccctactgcg gctgggacca gggccgctgc
2101 atctccatct acagctccga acggtcagtg ctgcaatcca ttaatccagc
2151 cgagccacac aaggagtgtc ccaaccccaa accagacaag gccccactgc
2201 agaaggtttc cctggcccca aactctcgct actacctgag ctgccccatg
2251 gaatcccgcc acgccaccta ctcatggcgc cacaaggaga acgtggagca
2301 gagctgcgaa cctggtcacc agagccccaa ctgcatcctg ttcatcgaga
2351 acctcacggc gcagcagtac ggccactact tctgcgaggc ccaggagggc
2401 tcctacttcc gcgaggctca gcactggcag ctgctgcccg aggacggcat
2451 catggccgag cacctgctgg gtcatgcctg tgccctggct gcctccctct
2501 ggctgggggt gctgcccaca ctcactcttg gcttgctggt ccacgtgaag
2551 cttGGGCCCG AACAAAAACT CATCTCAGAA GAGGATCTGA ATAGCGCCGT
2601 CGACCATCAT CATCATCATC ATTGAGTTTA TCCAGCACAG TGGCGGCCGC
2651 TCGAGTCTAG AGGGCCCGTT TAAACCCGCT GATCAGCCTC GACTGTGCCT
2701 TCTAGTTGCC AGCCATCTGT TGTTTGCCCC TCCCCCGTGC CTTCCTTGAC
2751 CCTGGAAGGT GCCACTCCCA CTGTCCTTTC CTAATAAAAT GAGGAAATTG
2801 CATCGCATTG TCTGAGTAGG TGTCATTCTA TTCTGGGGGG TGGGGTGGGG
2851 CAGGACAGCA AGGGGGAGGA TTGGGAAGAC AATAGCAGGC ATGCTGGGGA
2901 TGCGGTGGGC TCTATGGCTT CTGAGGCGGA AAGAACCAGC TGGGGCTCTA
2951 GGGGGTATCC CCACGCGCCC TGTAGCGGCG CATTAAGCGC GGCGGGTGTG
3001 GTGGTTACGC GCAGCGTGAC CGCTACACTT GCCAGCGCCC TAGCGCCCGC
3051 TCCTTTCGCT TTCTTCCCTT CCTTTCTCGC CACGTTCGCC GGCTTTCCCC

3101 GTCAAGCTCT AAATCGGGGC ATCCCTTTAG GGTTCCGATT TAGTGCTTTA
3151 CGGCACCTCG ACCCCAAAAA ACTTGATTAG GGTGATGGTT CACGTAGTGG
3201 GCCATCGCCC TGATAGACGG TTTTTCGCCC TTTGACGTTG GAGTCCACGT
3251 TCTTTAATAG TGGACTCTTG TTCCAAACTG GAACAACACT CAACCCTATC
3301 TCGGTCTATT CTTTTGATTT ATAAGGGATT TTGGGGATTT CGGCCTATTG
3351 GTTAAAAAAT GAGCTGATTT AACAAAAATT TAACGCGAAT TAATTCTGTG
3401 GAATGTGTGT CAGTTAGGGT GTGGAAAGTC CCCAGGCTCC CCAGGCAGGC
3451 AGAAGTATGC AAAGCATGCA TCTCAATTAG TCAGCAACCA GGTGTGGAAA
3501 GTCCCCAGGC TCCCCAGCAG GCAGAAGTAT GCAAAGCATG CATCTCAATT
3551 AGTCAGCAAC CATAGTCCCG CCCCTAACTC CGCCCATCCC GCCCCTAACT
3601 CCGCCCAGTT CCGCCCATTC TCCGCCCCAT GGCTGACTAA TTTTTTTTAT
3651 TTATGCAGAG GCCGAGGCCG CCTCTGCCTC TGAGCTATTC CAGAAGTAGT
3701 GAGGAGGCTT TTTTGGAGGC CTAGGCTTTT GCAAAAAGCT CCCGGGAGCT
3751 TGTATATCCA TTTTCGGATC TGATCAAGAG ACAGGATGAG GATCGTTTCG
3801 CATGATTGAA CAAGATGGAT TGCACGCAGG TTCTCCGGCC GCTTGGGTGG
3851 AGAGGCTATT CGGCTATGAC TGGGCACAAC AGACAATCGG CTGCTCTGAT
3901 GCCGCCGTGT TCCGGCTGTC AGCGCAGGGG CGCCCGGTTC TTTTTGTCAA
3951 GACCGACCTG TCCGGTGCCC TGAATGAACT GCAGGACGAG GCAGCGCGGC
4001 TATCGTGGCT GGCCACGACG GGCGTTCCTT GCGCAGCTGT GCTCGACGTT
4051 GTCACTGAAG CGGGAAGGGA CTGGCTGCTA TTGGGCGAAG TGCCGGGGCA
4101 GGATCTCCTG TCATCTCACC TTGCTCCTGC CGAGAAAGTA TCCATCATGG
4151 CTGATGCAAT GCGGCGGCTG CATACGCTTG ATCCGGCTAC CTGCCCATTC
4201 GACCACCAAG CGAAACATCG CATCGAGCGA GCACGTACTC GGATGGAAGC
4251 CGGTCTTGTC GATCAGGATG ATCTGGACGA AGAGCATCAG GGGCTCGCGC
4301 CAGCCGAACT GTTCGCCAGG CTCAAGGCGC GCATGCCCGA CGGCGAGGAT
4351 CTCGTCGTGA CCCATGGCGA TGCCTGCTTG CCGAATATCA TGGTGGAAAA
4401 TGGCCGCTTT TCTGGATTCA TCGACTGTGG CCGGCTGGGT GTGGCGGACC
4451 GCTATCAGGA CATAGCGTTG GCTACCCGTG ATATTGCTGA AGAGCTTGGC
4501 GGCGAATGGG CTGACCGCTT CCTCGTGCTT TACGGTATCG CCGCTCCCGA
4551 TTCGCAGCGC ATCGCCTTCT ATCGCCTTCT TGACGAGTTC TTCTGAGCGG
4601 GACTCTGGGG TTCGAAATGA CCGACCAAGC GACGCCCAAC CTGCCATCAC
4651 GAGATTTCGA TTCCACCGCC GCCTTCTATG AAAGGTTGGG CTTCGGAATC
4701 GTTTTCCGGG ACGCCGGCTG GATGATCCTC CAGCGCGGGG ATCTCATGCT
4751 GGAGTTCTTC GCCCACCCCA ACTTGTTTAT TGCAGCTTAT AATGGTTACA
4801 AATAAAGCAA TAGCATCACA AATTTCACAA ATAAAGCATT TTTTTCACTG

4851 CATTCTAGTT GTGGTTTGTC CAAACTCATC AATGTATCTT ATCATGTCTG
4901 TATACCGTCG ACCTCTAGCT AGAGCTTGGC GTAATCATGG TCATAGCTGT
4951 TTCCTGTGTG AAATTGTTAT CCGCTCACAA TTCCACACAA CATACGAGCC
5001 GGAAGCATAA AGTGTAAAGC CTGGGGTGCC TAATGAGTGA GCTAACTCAC
5051 ATTAATTGCG TTGCGCTCAC TGCCCGCTTT CCAGTCGGGA AACCTGTCGT
5101 GCCAGCTGCA TTAATGAATC GGCCAACGCG CGGGGAGAGG CGGTTTGCGT
5151 ATTGGGCGCT CTTCCGCTTC CTCGCTCACT GACTCGCTGC GCTCGGTCGT
5201 TCGGCTGCGG CGAGCGGTAT CAGCTCACTC AAAGGCGGTA ATACGGTTAT
5251 CCACAGAATC AGGGGATAAC GCAGGAAAGA ACATGTGAGC AAAAGGCCAG
5301 CAAAAGGCCA GGAACCGTAA AAAGGCCGCG TTGCTGGCGT TTTTCCATAG
5351 GCTCCGCCCC CCTGACGAGC ATCACAAAAA TCGACGCTCA AGTCAGAGGT
5401 GGCGAAACCC GACAGGACTA TAAAGATACC AGGCGTTTCC CCCTGGAAGC
5451 TCCCTCGTGC GCTCTCCTGT TCCGACCCTG CCGCTTACCG GATACCTGTC
5501 CGCCTTTCTC CCTTCGGGAA GCGTGGCGCT TTCTCAATGC TCACGCTGTA
5551 GGTATCTCAG TTCGGTGTAG GTCGTTCGCT CCAAGCTGGG CTGTGTGCAC
5601 GAACCCCCCG TTCAGCCCGA CCGCTGCGCC TTATCCGGTA ACTATCGTCT
5651 TGAGTCCAAC CCGGTAAGAC ACGACTTATC GCCACTGGCA GCAGCCACTG
5701 GTAACAGGAT TAGCAGAGCG AGGTATGTAG GCGGTGCTAC AGAGTTCTTG
5751 AAGTGGTGGC CTAACTACGG CTACACTAGA AGGACAGTAT TTGGTATCTG
5801 CGCTCTGCTG AAGCCAGTTA CCTTCGGAAA AAGAGTTGGT AGCTCTTGAT
5851 CCGGCAAACA AACCACCGCT GGTAGCGGTG GTTTTTTTGT TTGCAAGCAG
5901 CAGATTACGC GCAGAAAAAA AGGATCTCAA GAAGATCCTT TGATCTTTTC
5951 TACGGGGTCT GACGCTCAGT GGAACGAAAA CTCACGTTAA GGGATTTTGG
6001 TCATGAGATT ATCAAAAAGG ATCTTCACCT AGATCCTTTT AAATTAAAAA
6051 TGAAGTTTTA AATCAATCTA AAGTATATAT GAGTAAACTT GGTCTGACAG
6101 TTACCAATGC TTAATCAGTG AGGCACCTAT CTCAGCGATC TGTCTATTTC
6151 GTTCATCCAT AGTTGCCTGA CTCCCCGTCG TGTAGATAAC TACGATACGG
6201 GAGGGCTTAC CATCTGGCCC CAGTGCTGCA ATGATACCGC GAGACCCACG
6251 CTCACCGGCT CCAGATTTAT CAGCAATAAA CCAGCCAGCC GGAAGGGCCG
6301 AGCGCAGAAG TGGTCCTGCA ACTTTATCCG CCTCCATCCA GTCTATTAAT
6351 TGTTGCCGGG AAGCTAGAGT AAGTAGTTCG CCAGTTAATA GTTTGCGCAA
6401 CGTTGTTGCC ATTGCTACAG GCATCGTGGT GTCACGCTCG TCGTTTGGTA
6451 TGGCTTCATT CAGCTCCGGT TCCCAACGAT CAAGGCGAGT TACATGATCC
6501 CCCATGTTGT GCAAAAAAGC GGTTAGCTCC TTCGGTCCTC CGATCGTTGT
6551 CAGAAGTAAG TTGGCCGCAG TGTTATCACT CATGGTTATG GCAGCACTGC

6601 ATAATTCTCT TACTGTCATG CCATCCGTAA GATGCTTTTC TGTGACTGGT
6651 GAGTACTCAA CCAAGTCATT CTGAGAATAG TGTATGCGGC GACCGAGTTG
6701 CTCTTGCCCG GCGTCAATAC GGGATAATAC CGCGCCACAT AGCAGAACTT
6751 TAAAAGTGCT CATCATTGGA AAACGTTCTT CGGGGCGAAA ACTCTCAAGG
6801 ATCTTACCGC TGTTGAGATC CAGTTCGATG TAACCCACTC GTGCACCCAA
6851 CTGATCTTCA GCATCTTTTA CTTTCACCAG CGTTTCTGGG TGAGCAAAAA
6901 CAGGAAGGCA AAATGCCGCA AAAAAGGGAA TAAGGGCGAC ACGGAAATGT
6951 TGAATACTCA TACTCTTCCT TTTTCAATAT TATTGAAGCA TTTATCAGGG
7001 TTATTGTCTC ATGAGCGGAT ACATATTTGA ATGTATTTAG AAAAATAAAC
7051 AAATAGGGGT TCCGCGCACA TTTCCCCGAA AAGTGCCACC TGACGTCGAC
7101 GGATCGGG

Table12: Sequence of the recombinant plasmid pQE30-H-SemaL-BH (SEQ ID NO.:39)

1 CTCGAGAAAT CATAAAAAAT TTATTTGCTT TGTGAGCGGA TAACAATTAT
51 AATAGATTCA ATTGTGAGCG GATAACAATT TCACACAGAA TTCATTAAAG
101 AGGAGAAATT AACTATGAGA GGATCGCATC ACCATCACCA TCACGGAtcc
151 ctggttctgt ttgaagggga cgaggtgtat tccaccatcc ggaagcagga
201 atacaatggg aagatccctc ggttccgccg catccggggc gagagtgagc
251 tgtacaccag tgatactgtc atgcagaacc cacagttcat caaagccacc
301 atcgtgcacc aagaccaggc ttacgatgac aagatctact acttcttccg
351 agaggacaat cctgacaaga atcctgaggc tcctctcaat gtgtcccgtg
401 tggcccagtt gtgcaggggg gaccagggtg ggaaagttc actgtcagtc
451 tccaagtgga acacttttct gaaagccatg ctggtatgca gtgatgctgc
501 caccaacaag aacttcaaca ggctgcaaga cgtcttcctg ctccctgacc
551 ccagcggcca gtggagggac accagggtct atggtgtttt ctccaacccc
601 tggaactact cagccgtctg tgtgtattcc ctcggtgaca ttgacaaggt
651 cttccgtacc tcctcactca agggctacca ctcaagcctt cccaacccgc
701 ggcctggcaa gtgcctccca gaccagcagc cgatacccac agaAAGCTTA
751 ATTAGCTGAG CTTGGACTCC TGTTGATAGA TCCAGTAATG ACCTCAGAAC
801 TCCATCTGGA TTTGTTCAGA ACGCTCGGTT GCCGCCGGGC GTTTTTTATT
851 GGTGAGAATC CAAGCTAGCT TGGCGAGATT TTCAGGAGCT AAGGAAGCTA
901 AAATGGAGAA AAAAATCACT GGATATACCA CCGTTGATAT ATCCCAATGG

951 CATCGTAAAG AACATTTTGA GGCATTTCAG TCAGTTGCTC AATGTACCTA
1001 TAACCAGACC GTTCAGCTGG ATATTACGGC CTTTTTAAAG ACCGTAAAGA
1051 AAAATAAGCA CAAGTTTTAT CCGGCCTTTA TTCACATTCT TGCCCGCCTG
1101 ATGAATGCTC ATCCGGAATT TCGTATGGCA ATGAAAGACG GTGAGCTGGT
1151 GATATGGGAT AGTGTTCACC CTTGTTACAC CGTTTTCCAT GAGCAAACTG
1201 AAACGTTTTC ATCGCTCTGG AGTGAATACC ACGACGATTT CCGGCAGTTT
1251 CTACACATAT ATTCGCAAGA TGTGGCGTGT TACGGTGAAA ACCTGGCCTA
1301 TTTCCCTAAA GGGTTTATTG AGAATATGTT TTTCGTCTCA GCCAATCCCT
1351 GGGTGAGTTT CACCAGTTTT GATTTAAACG TGGCCAATAT GGACAACTTC
1401 TTCGCCCCCG TTTTCACCAT GGGCAAATAT TATACGCAAG GCGACAAGGT
1451 GCTGATGCCG CTGGCGATTC AGGTTCATCA TGCCGTCTGT GATGGCTTCC
1501 ATGTCGGCAG AATGCTTAAT GAATTACAAC AGTACTGCGA TGAGTGGCAG
1551 GGCGGGGCGT AATTTTTTTA AGGCAGTTAT TGGTGCCCTT AAACGCCTGG
1601 GGTAATGACT CTCTAGCTTG AGGCATCAAA TAAAACGAAA GGCTCAGTCG
1651 AAAGACTGGG CCTTTCGTTT TATCTGTTGT TTGTCGGTGA ACGCTCTCCT
1701 GAGTAGGACA AATCCGCCGC TCTAGAGCTG CCTCGCGCGT TTCGGTGATG
1751 ACGGTGAAAA CCTCTGACAC ATGCAGCTCC CGGAGACGGT CACAGCTTGT
1801 CTGTAAGCGG ATGCCGGGAG CAGACAAGCC CGTCAGGGCG CGTCAGCGGG
1851 TGTTGGCGGG TGTCGGGGCG CAGCCATGAC CCAGTCACGT AGCGATAGCG
1901 GAGTGTATAC TGGCTTAACT ATGCGGCATC AGAGCAGATT GTACTGAGAG
1951 TGCACCATAT GCGGTGTGAA ATACCGCACA GATGCGTAAG GAGAAAATAC
2001 CGCATCAGGC GCTCTTCCGC TTCCTCGCTC ACTGACTCGC TGCGCTCGGT
2051 CTGTCGGCTG CGGCGAGCGG TATCAGCTCA CTCAAAGGCG GTAATACGGT
2101 TATCCACAGA ATCAGGGGAT AACGCAGGAA AGAACATGTG AGCAAAAGGC
2151 CAGCAAAAGG CCAGGAACCG TAAAAAGGCC GCGTTGCTGG CGTTTTTCCA
2201 TAGGCTCCGC CCCCCTGACG AGCATCACAA AAATCGACGC TCAAGTCAGA
2251 GGTGGCGAAA CCCGACAGGA CTATAAAGAT ACCAGGCGTT TCCCCCTGGA
2301 AGCTCCCTCG TGCGCTCTCC TGTTCCGACC CTGCCGCTTA CCGGATACCT
2351 GTCCGCCTTT CTCCCTTCGG GAAGCGTGGC GCTTTCTCAA TGCTCACGCT
2401 GTAGGTATCT CAGTTCGGTG TAGGTCGTTC GCTCCAAGCT GGGCTGTGTG
2451 CACGAACCCC CCGTTCAGCC CGACCGCTGC GCCTTATCCG GTAACTATCG
2501 TCTTGAGTCC AACCCGGTAA GACACGACTT ATCGCCACTG GCAGCAGCCA
2551 CTGGTAACAG GATTAGCAGA GCGAGGTATG TAGGCGGTGC TACAGAGTTC
2601 TTGAAGTGGT GGCCTAACTA CGGCTACACT AGAAGGACAG TATTTGGTAT
2651 CTGCGCTCTG CTGAAGCCAG TTACCTTCGG AAAAAGAGTT GGTAGCTCTT

2701 GATCCGGCAA ACAAACCACC GCTGGTAGCG GTGGTTTTTT TGTTTGCAAG
2751 CAGCAGATTA CGCGCAGAAA AAAAGGATCT CAAGAAGATC CTTTGATCTT
2801 TTCTACGGGG TCTGACGCTC AGTGGAACGA AAACTCACGT TAAGGGATTT
2851 TGGTCATGAG ATTATCAAAA AGGATCTTCA CCTAGATCCT TTTAAATTAA
2901 AAATGAAGTT TTAAATCAAT CTAAAGTATA TATGAGTAAA CTTGGTCTGA
2951 CAGTTACCAA TGCTTAATCA GTGAGGCACC TATCTCAGCG ATCTGTCTAT
3001 TTCGTTCATC CATAGCTGCC TGACTCCCCG TCGTGTAGAT AACTACGATA
3051 CGGGAGGGCT TACCATCTGG CCCCAGTGCT GCAATGATAC CGCGAGACCC
3101 ACGCTCACCG GCTCCAGATT TATCAGCAAT AAACCAGCCA GCCGGAAGGG
3151 CCGAGCGCAG AAGTGGTCCT GCAACTTTAT CCGCCTCCAT CCAGTCTATT
3201 AATTGTTGCC GGGAAGCTAG AGTAAGTAGT TCGCCAGTTA ATAGTTTGCG
3251 CAACGTTGTT GCCATTGCTA CAGGCATCGT GGTGTCACGC TCGTCGTTTG
3301 GTATGGCTTC ATTCAGCTCC GGTTCCCAAC GATCAAGGCG AGTTACATGA
3351 TCCCCCATGT TGTGCAAAAA AGCGGTTAGC TCCTTCGGTC CTCCGATCGT
3401 TGTCAGAAGT AAGTTGGCCG CAGTGTTATC ACTCATGGTT ATGGCAGCAC
3451 TGCATAATTC TCTTACTGTC ATGCCATCCG TAAGATGCTT TTCTGTGACT
3501 GGTGAGTACT CAACCAAGTC ATTCTGAGAA TAGTGTATGC GGCGACCGAG
3551 TTGCTCTTGC CCGGCGTCAA TACGGGATAA TACCGCGCCA CATAGCAGAA
3601 CTTTAAAAGT GCTCATCATT GGAAAACGTT CTTCGGGGCG AAAACTCTCA
3651 AGGATCTTAC CGCTGTTGAG ATCCAGTTCG ATGTAACCCA CTCGTGCACC
3701 CAACTGATCT TCAGCATCTT TTACTTTCAC CAGCGTTTCT GGGTGAGCAA
3751 AAACAGGAAG GCAAAATGCC GCAAAAAAGG GAATAAGGGC GACACGGAAA
3801 TGTTGAATAC TCATACTCTT CCTTTTTCAA TATTATTGAA GCATTTATCA
3851 GGGTTATTGT CTCATGAGCG GATACATATT TGAATGTATT TAGAAAAATA
3901 AACAAATAGG GGTTCCGCGC ACATTTCCCC GAAAAGTGCC ACCTGACGTC
3951 TAAGAAACCA TTATTATCAT GACATTAACC TATAAAAATA GGCGTATCAC
4001 GAGGCCCTTT CGTCTTCAC

Table13: Sequence of the recombinant plasmid pQE31-H-SemaL-SH (SEQ ID NO.: 40)

1 CTCGAGAAAT CATAAAAAAT TTATTTGCTT TGTGAGCGGA TAACAATTAT
51 AATAGATTCA ATTGTGAGCG GATAACAATT TCACACAGAA TTCATTAAAG
101 AGGAGAAATT AACTATGAGA GGATCGCATC ACCATCACCA TCACACGGAT

151 CCGCATGCga gctcccagtg ggaggtgagc caggtgcccc tggacctgtg

201 tgaggtctat ggcgggggct gccacggttg cctcatgtcc cgagacccct

251 actgcggctg ggaccagggc cgctgcatct ccatctacag ctccgaacgg

301 tcagtgctgc aatccattaa tccagccgag ccacacaagg agtgtcccaa

351 ccccaaacca gacaaggccc cactgcagaa ggtttccctg gccccaaact

401 ctcgctacta cctgagctgc cccatggaat cccgccacgc cacctactca

451 tggcgccaca aggagaacgt ggagcagagc tgcgaacctg gtcaccagag

501 ccccaactgc atcctgttca tcgagaacct cacggcgcag cagtacggcc

551 actacttctg cgaggcccag gagggctcct acttccgcga ggctcagcac

601 tggcagctgc tgcccgagga cggcatcatg gccgagcacc tgctgggtca

651 tgcctgtgcc ctggctgcct ccctctggct gggggtgctg cccacactca

701 ctcttggctt gctggtccac gtgaagcttA ATTAGCTGAG CTTGGACTCC

751 TGTTGATAGA TCCAGTAATG ACCTCAGAAC TCCATCTGGA TTTGTTCAGA

801 ACGCTCGGTT GCCGCCGGGC GTTTTTTATT GGTGAGAATC CAAGCTAGCT

851 TGGCGAGATT TTCAGGAGCT AAGGAAGCTA AAATGGAGAA AAAAATCACT

901 GGATATACCA CCGTTGATAT ATCCCAATGG CATCGTAAAG AACATTTTGA

951 GGCATTTCAG TCAGTTGCTC AATGTACCTA TAACCAGACC GTTCAGCTGG

1001 ATATTACGGC CTTTTTAAAG ACCGTAAAGA AAAATAAGCA CAAGTTTTAT

1051 CCGGCCTTTA TTCACATTCT TGCCCGCCTG ATGAATGCTC ATCCGGAATT

1101 TCGTATGGCA ATGAAAGACG GTGAGCTGGT GATATGGGAT AGTGTTCACC

1151 CTTGTTACAC CGTTTTCCAT GAGCAAACTG AAACGTTTTC ATCGCTCTGG

1201 AGTGAATACC ACGACGATTT CCGGCAGTTT CTACACATAT ATTCGCAAGA

1251 TGTGGCGTGT TACGGTGAAA ACCTGGCCTA TTTCCCTAAA GGGTTTATTG

1301 AGAATATGTT TTTCGTCTCA GCCAATCCCT GGGTGAGTTT CACCAGTTTT

1351 GATTTAAACG TGGCCAATAT GGACAACTTC TTCGCCCCCG TTTTCACCAT

1401 GGGCAAATAT TATACGCAAG GCGACAAGGT GCTGATGCCG CTGGCGATTC

1451 AGGTTCATCA TGCCGTCTGT GATGGCTTCC ATGTCGGCAG AATGCTTAAT

1501 GAATTACAAC AGTACTGCGA TGAGTGGCAG GGCGGGGCGT AATTTTTTTA

1551 AGGCAGTTAT TGGTGCCCTT AAACGCCTGG GGTAATGACT CTCTAGCTTG

1601 AGGCATCAAA TAAAACGAAA GGCTCAGTCG AAAGACTGGG CCTTTCGTTT

1651 TATCTGTTGT TTGTCGGTGA ACGCTCTCCT GAGTAGGACA AATCCGCCGC

1701 TCTAGAGCTG CCTCGCGCGT TTCGGTGATG ACGGTGAAAA CCTCTGACAC

1751 ATGCAGCTCC CGGAGACGGT CACAGCTTGT CTGTAAGCGG ATGCCGGGAG

1801 CAGACAAGCC CGTCAGGGCG CGTCAGCGGG TGTTGGCGGG TGTCGGGGCG

1851 CAGCCATGAC CCAGTCACGT AGCGATAGCG GAGTGTATAC TGGCTTAACT

1901 ATGCGGCATC AGAGCAGATT GTACTGAGAG TGCACCATAT GCGGTGTGAA
1951 ATACCGCACA GATGCGTAAG GAGAAAATAC CGCATCAGGC GCTCTTCCGC
2001 TTCCTCGCTC ACTGACTCGC TGCGCTCGGT CTGTCGGCTG CGGCGAGCGG
2051 TATCAGCTCA CTCAAAGGCG GTAATACGGT TATCCACAGA ATCAGGGGAT
2101 AACGCAGGAA AGAACATGTG AGCAAAAGGC CAGCAAAAGG CCAGGAACCG
2151 TAAAAAGGCC GCGTTGCTGG CGTTTTTCCA TAGGCTCCGC CCCCCTGACG
2201 AGCATCACAA AAATCGACGC TCAAGTCAGA GGTGGCGAAA CCCGACAGGA
2251 CTATAAAGAT ACCAGGCGTT TCCCCCTGGA AGCTCCCTCG TGCGCTCTCC
2301 TGTTCCGACC CTGCCGCTTA CCGGATACCT GTCCGCCTTT CTCCCTTCGG
2351 GAAGCGTGGC GCTTTCTCAA TGCTCACGCT GTAGGTATCT CAGTTCGGTG
2401 TAGGTCGTTC GCTCCAAGCT GGGCTGTGTG CACGAACCCC CCGTTCAGCC
2451 CGACCGCTGC GCCTTATCCG GTAACTATCG TCTTGAGTCC AACCCGGTAA
2501 GACACGACTT ATCGCCACTG GCAGCAGCCA CTGGTAACAG GATTAGCAGA
2551 GCGAGGTATG TAGGCGGTGC TACAGAGTTC TTGAAGTGGT GGCCTAACTA
2601 CGGCTACACT AGAAGGACAG TATTTGGTAT CTGCGCTCTG CTGAAGCCAG
2651 TTACCTTCGG AAAAAGAGTT GGTAGCTCTT GATCCGGCAA ACAAACCACC
2701 GCTGGTAGCG GTGGTTTTTT TGTTTGCAAG CAGCAGATTA CGCGCAGAAA
2751 AAAAGGATCT CAAGAAGATC CTTTGATCTT TTCTACGGGG TCTGACGCTC
2801 AGTGGAACGA AAACTCACGT TAAGGGATTT TGGTCATGAG ATTATCAAAA
2851 AGGATCTTCA CCTAGATCCT TTTAAATTAA AAATGAAGTT TTAAATCAAT
2901 CTAAAGTATA TATGAGTAAA CTTGGTCTGA CAGTTACCAA TGCTTAATCA
2951 GTGAGGCACC TATCTCAGCG ATCTGTCTAT TTCGTTCATC CATAGCTGCC
3001 TGACTCCCCG TCGTGTAGAT AACTACGATA CGGGAGGGCT TACCATCTGG
3051 CCCCAGTGCT GCAATGATAC CGCGAGACCC ACGCTCACCG GCTCCAGATT
3101 TATCAGCAAT AAACCAGCCA GCCGGAAGGG CCGAGCGCAG AAGTGGTCCT
3151 GCAACTTTAT CCGCCTCCAT CCAGTCTATT AATTGTTGCC GGGAAGCTAG
3201 AGTAAGTAGT TCGCCAGTTA ATAGTTTGCG CAACGTTGTT GCCATTGCTA
3251 CAGGCATCGT GGTGTCACGC TCGTCGTTTG GTATGGCTTC ATTCAGCTCC
3301 GGTTCCCAAC GATCAAGGCG AGTTACATGA TCCCCCATGT TGTGCAAAAA
3351 AGCGGTTAGC TCCTTCGGTC CTCCGATCGT TGTCAGAAGT AAGTTGGCCG
3401 CAGTGTTATC ACTCATGGTT ATGGCAGCAC TGCATAATTC TCTTACTGTC
3451 ATGCCATCCG TAAGATGCTT TTCTGTGACT GGTGAGTACT CAACCAAGTC
3501 ATTCTGAGAA TAGTGTATGC GGCGACCGAG TTGCTCTTGC CCGGCGTCAA
3551 TACGGGATAA TACCGCGCCA CATAGCAGAA CTTTAAAAGT GCTCATCATT
3601 GGAAAACGTT CTTCGGGGCG AAAACTCTCA AGGATCTTAC CGCTGTTGAG

3651 ATCCAGTTCG ATGTAACCCA CTCGTGCACC CAACTGATCT TCAGCATCTT

3701 TTACTTTCAC CAGCGTTTCT GGGTGAGCAA AAACAGGAAG GCAAAATGCC

3751 GCAAAAAAGG GAATAAGGGC GACACGGAAA TGTTGAATAC TCATACTCTT

3801 CCTTTTTCAA TATTATTGAA GCATTTATCA GGGTTATTGT CTCATGAGCG

3851 GATACATATT TGAATGTATT TAGAAAAATA AACAAATAGG GGTTCCGCGC

3901 ACATTTCCCC GAAAAGTGCC ACCTGACGTC TAAGAAACCA TTATTATCAT

3951 GACATTAACC TATAAAAATA GGCGTATCAC GAGGCCCTTT CGTCTTCAC

Table14: (Partial) nucleotide sequence of the human semaphorin L gene. (8888 nucleotides) (SEQ ID NO.: 41):

GAGCCGCACACGGTGCTTTTCCACGAGCCAGGCAGCTCCTCTGTGTGGGTGGGAGGACGT

GGCAAGGTCTACCTCTTTGACTTCCCCGAGGGCAAGAACGCATCTGTGCGCACGGTGAGC

CTCTCTCTTCCCCCAACACCCCCCCTACCCTCTTATCTCCCCTCTGGCCCTGCCAAGGGT

CCTCAGGGAATCCGAGGGAGCTGGCTTCTCTTCCTAAACTGCCCCCACCTCCGTATCCTA

TAAATGGCTCCTGGGGGAGGCTCCCTAAAGGTAGTCCAGATTGGAGTGGGGAGCTGGGGC

GGTGTGGAGAAAAACAGGAGCTAATGGGCCTGGCCAGCTGGGCAGCGCTGCTGCGGAAAG

CCCAGGCTGGAAGCTGGGCCCCAGAGCCCATGCCTGGTCTTCTGAACCCTCTGGGCCTCA

GCTCTGGATATGAGACCCTGTTTGACCTCAGGTAGATCACTCACCCTCTCAGAGCCCCAG

TTGCTCATCTGTCAGATGAGAATAATGGTTGCTTCCTTTGGGGCTTATCCTGAGGCTGTG

TGGAAAGCATTTCAGGGGTACCTCACCCCTGGCAGATTGAACTAATGCTTCTCCCCTTCC

CCAGGTGAATATCGGCTCCACAAAGGGGTCCTGTCTGGATAAGCGGGTGAGCGGGGGAGG

GATCTGGAGGGGTCTGAGCCACTTGGTAAAGGGAGAGGAGACCCTGAGGGTCTAAGGAAG

GAAGCATGGCCCTGCCCCACGAGTCCCAGACTGATGGGGAGACGTGGTCCTCTGTGCTTA

GGGGATGGCGTCAGCTGCACACACTCTGGGCTGTCCCGGGAGGCTGTCACCTATGCTAAG

CCCTTCTGACACCTTCTTCCCTGATCCTGGGGGTCCTAGTGCTAGGCTTGCCAGGGCCTT

CCAGCAACCAATTTCTCTCCTCCCTTCTCTCTTCCCCGGGCAGGACTGCGAGAACTACAT

CACTCTCCTGGAGAGGCGGAGTGAGGGGCTGCTGGCCTGTGGCACCAACGCCCGGCACCC

CAGCTGCTGGAACCTGGTGAGAAGGCTGCTCCCCATGTGCCTGATCAGCTCACCTTCTAC

TGCGTGGGCTTCTGCCCCTCATGGTGGGAAGGAGATGGCGAGACTCCAATGCTGGCCTTG

CCCTGGGAGGATGGGGCTCCTGGCCGAGAAACTGGCCGTCATGGGAGGCAGTGGCTGTGG

GATTATGTGGCCATCCAACCCTCTGGATCTCCCACAGGTGAATGGCACTGTGGTGCCACT

TGGCGAGATGAGAGGCTACGCCCCCTTCAGCCCGGACGAGAACTCCCTGGTTCTGTTTGA

AGGTTGGGGCATGCTTCGGAACTGGGCTGGGAGCAGGATGGTCAGCTCTTTGTCCAGTGT

CCGGAGGAGGGACTTCCAGGAGCTGCCTGCCCTTACTCATTTCTCCCTCCCACTGACCCC

AGGGGACGAGGTGTATTCCACCATCCGGAAGCAGGAATACAATGGGAAGATCCCTCGGTT

CCGCCGCATCCGGGGCGAGAGTGAGCTGTACACCAGTGATACTGTCATGCAGAGTGAGTC

AGGCTCCGGCTGGGCTGAGGGTGGGCAAGGGGGTGTGAGCACTTAAGGTGGCAGATGGGA

TCCTGATGTTTCTGGGAGGGCTCCCTGAGGGCCGCTGGGGCCATGCAGGAAAGCAGGACC

TTGGTATAGGCCTGAGAAGTTAGGGTTGGCTGGGAGCAGAGGAACAGACAAGGTATAGCA

GTGGGATGGGCCCAGCCCTCTTCAGGAACACAAACAGAGGGAGCCCCAGACCCAGTGCAG

GGTCCCCAGGAGCCAAAGTTTATCCTCTGCTGAGTTCACGTGGAGGCAGCCCCCCAACTC

CCTCCTCATCAGGGCTCTGCCAATTGAGCAGAAGTGACATAGGGGCCCCCAGGGACCTTC

CCCCACTCCCCAGGCATGAAGTCATTGCTCCTGGGCCGATGACATCTTTGTAGGAAGAGG

GCAAAACAGGTGTGGGGTGGAGGTGCAGGGTCTAGGGCCCCTCGGGGAGTTGGACCTGAT

GTTATGAGTCCTATTCCAGATCTGATTTGCCATGGTTTGTGCAGACCCGAAGGAGGGAGG

AGAGTGTGCAGGGTTGGAATGGTCTCCCGGGCAAGCTTCCCAGCCTTACGCCCATTCGCT

TCTGTGCCCTGGCAGACCCACAGTTCATCAAAGCCACCATCGTGCACCAAGACCAGGCTT

ACGATGACAAGATCTACTACTTCTTCCGAGAGGACAATCCTGACAAGAATCCTGAGGCTC

CTCTCAATGTGTCCCGTGTGGCCCAGTTGTGCAGGGTGAACACGGGCGTGAGGGCTGCTG

GCTACGTGTCTGTGCATGAATAGGCCTGAGTGAGGGTGAGTTCTGTGTGTCCGTGTGCAT

GTAGAAGTTGTGTGGATGTATGAGTGGGTCTGTGTCAGGGACTGTGGGAGCAGCTGTGTG

TGCATGGAGCATCATGTGTCTGTGTGTGGGTAAAGGTGGCTGAGCTCCTGTGCACGTATG

ATGGCGTGTGAGCGTGTGTATGATGGGGTGTGTGTGTGTGTGTGTGTGTGTTTTGCCT

GTGTGAATGTGCTGTGCCACGTATGTGGGTGCGTGAGTCAGTAAATGTGTGTCTGAGTCC

GTCTGCTCTGTGGGGACCTGGCACTCTCACCTGCCCTGACCCTGGGCACTGCTGGCCCTG

GGCTCTGGATCAGCCAGGCCTGCTTGCAGGAGTCTCATCTGGAGACCTGCCCTGAGTCCT

GGGGCACCCCCGGCAGGTCCTGGCCCCTCGCAGCCTGCCTTCCTCCTCTGGGCCCAGGTG

TTGATATTGCTGGCAGTGGTTTCCTGGGGTGTGTGGGGAAGCCCGGGCAGGTGCTGAGGG

GCCTCTTCTCCCCTCTACCCTTCCAGGGGGACCAGGGTGGGGAAAGTTCACTGTCAGTCT

CCAAGTGGAACACTTTTCTGAAAGCCATGCTGGTATGCAGTGATGCTGCCACCAACAAGA

ACTTCAACAGGCTGCAAGACGTCTTCCTGCTCCCTGACCCCAGCGGCCAGTGGAGGGACA

CCAGGGTCTATGGTGTTTTCTCCAACCCCTGGTGAGTGGCCCTTGTCCTGGGGCCGGGGC

TGGCATTGGTTCAGTGTCCAGTAGGGACAGGAGGCCTTGGGCCCTGCTGAGGGCCTCCCT

GGTGTGGCAGGAGCAGGGGCTGCAGGCTCAAGAGGCTGGGCTGTTGCTGGGTGTGGGGTG

GGGGGACAGCCAGTGCGATGTATGTACTGTTGTGTGAGTGAGTCTGCACTCATGGGTGTG

TGTGCATGCCCTATATGCACACTCATGACTGCACTTGTGCCTGTGTGTCCCACCACCTGC

TTGTGCCGAGAGTGGACACTGGGCCCAGGAGGAAGCTGCTGAAGCATCTCTCGGGGAGCT

GGGTGCTATTACACCTGCTCAGGCACTGCCTGAGCCCGATAATTCACACTTCTTAATCAC

TCTCATTGATTGAACACACGGCAGGCGGAAGTGTTGGGTGTGTGTGGGGAGAGTTAGGGA

TAGAGTGGAGGAAGCCAAGACCCTGCTCTGTGGCTCCTGGGTGAGTGGGTCCCCCAGGCT

GGGAAGGGGTTGGGGGTCTGGCCTCCTGGGGCATCAGCACCCCACAGCCTGTGCCCAGGG

AGGGCTAGAGAACTGCTCAGCCTATGATGGGGTTCCTCCTGCCTTGGGGTTGGGTAGAGC

AGATGGCCTCTAGACTCAGTGATTCTGTAACAGGATACAAGTTTGTGGTTTTAAATTGCA

GCACAAAGAAATTAGGCTGAACTCCTCTCCTTCCTCCTCTCCATCCCTCCCCATTTTCAG

TGGTGGTTGGCAACTCAGTGCCAGGCACAAGGCTGGCCTGGGTGAGTGGAGGTGGATGGG

TGGGTTCTGGGCCCCCCATTGAGCTGGTCTCCATGTCACTGCAGGAACTACTCAGCCGTC

TGTGTGTATTCCCTCGGTGACATTGACAAGGTCTTCCGTACCTCCTCACTCAAGGGCTAC

CACTCAAGCCTTCCCAACCCGCGGCCTGGCAAGGTGAGCGTGACACCAGCCGTGGCCCAG

GCCCAGCCCTCCTTCTGCCTCACCTCCCACCACCCCACTGACCTGGGCCTGCTCTCCTTG

CCCAGTGCCTCCCAGACCAGCAGCCGATACCCACAGAGACCTTCCAGGTGGCTGACCGTC

ACCCAGAGGTGGCGCAGAGGGTGGAGCCCATGGGGCCTCTGAAGACGCCATTGTTCCACT

CTAAATACCACTACCAGAAAGTGGCCGTCCACCGCATGCAAGCCAGCCACGGGGAGACCT

TTCATGTGCTTTACCTAACTACAGGTGAGAGGCTACCCCGGGACCCTCAGTTTGCTTTGT

AAAAACGGGCATGAAAGGTGTAAGGAATAATGTAGTTAACATCTGGTTGGATCTTTACAT

GTGGAAGGAATAATTGAGTGACTGGAGTTGTCAGGGGTTAATGTGTGTGGGTGTGGAAGA

GCCAGGCAGGGAGAGCTTCCTGGAGGAGGTAGGGGCAAGAGGGAAAGGGGGATGGGAGAA

AAGCAAGCACTGGGATTTGGAGGCGGAAATCTGGAGAGTCTGAGCAAAGCCAGGTGCACC

TTTGGTCCAGATGTCTGACTCAGGGAAGAAGATGGTAGGAAGAGACGTGGCAAATGAGGA

GGAGGGGCCTGAACCACAGGGATACTGGCCTCTGCCAGGCAGAATGAGGGAGTCAGGCCC

TGCGCCTGTCTTTGGGATTGTGCAGGTGAGAAGAAACATTTGAGGAGTTGATGGGGCACA

AATTAGGTATGGGGAAGGAGTTCCAGGGGGCAGAACCTTTGCCATCTCACAGAGGACAGG

GGCAGCTTCTCTTCTTCCCTGGAGTAGGCCCTGCTGGGGGAAGCTGGGTGGAATGCCGTG

GGAGATGCTCCTGCTTTCTGGAAAGCCACAGGACACGGAGGAGCCAGTCCTGAGTTGGGT

TTGTCGCAGCTTCCCATGCCAGCTGCCTTCCTTGAGACTGGAAAGGGCCTCTAGCACCCC

TGGGGCCATTCAATTCAGGCCCAGGCGCCCAACCTCAGTTGTTCACATTCCCCATGTGAT

CTCCTGTTGCTGCTTCACCTTGGGACTGTCTCGGCTTTGGTGACCTTGTAGGAAACTGGA

ACCCCAGCACCATTGTTTGGCTCCTGGAAGCCTTGGGGAGAGGAATTTCCCACAGGGCAG

GGCCTGGGTCCTGATTCCCTGCCTCTTTACTCCCTATTCATCCCGGCTACACCCTTGGGC

CCCCATCCTTGCTTGGCTCCAGTACTGGCTGGCACAGCTGTTGTGGTCATCCAGGGATGG

CAGGGCACTGGGGAACAGAAGAGAGAGGTCACACAGTGCGGAACTGGGAGCAGGAGCTAG

GACAAGGAAGGCTGGACTTGGGCCATGGATTCCCTTCCTGCAGACTTGGGAAGTGAGCAC

ACTTGAGTGATTAGAGAAGGTGTCTTCGTTCTAAGGGCAGTGGAGGAGGCACCATTTTGG

AGCCTGCATCATTCGTATTTGGGCTAGATTGAAAAATAGAGCTTTCTAAGTCCTCTGCAG

AGAATGGGAGGCTCTCACAACTGGGAGAAGTATTGGCTCTTTTCCTGAGAATTTTGCCAA
GGGTATGCTGTTACTGGGGCTGGTTTGGAAGGAGTATAGGGCATTATGTCTGTGAAGGCA
GTGGCTGGGGTGGGGCCTTATCAGGCCCAAGGAGCATCTGGCCACATCTCAGAGTCCACA
GATGAGGATCACGGATGTGTAGAGGAAACATCCTAGGCAGGCAATCATCTGACTGCTTTT
TTGGGGCAGGTGATGCCCTGGGAAATTGGGAGGGAGGGAGAGAGGGAGGTAGGCTATTCT
AGAAACTGGGAGAGCAGGTGAGGTAGGATTGGGAGGACCAGGGGTCAGGGTCCCCATTGG
TCCCTAATTGAGAACGGAGAGAGCATTGGTCTAGGAGGCAGGCAGCTCGGTTATAAGACC
TTGGGAACTCTTGATTTAGAATCCAAGATCCTTTTTAGATCTAGGATTTTATAAAATTAA
GATATCCCCTAAGATCAAATGCAACGTGGAGTCCTGAATTGGATCCTAGAACAGAAGAAG
GACATTTGTGGAAAAACTAGTGAAATCCAAATAAAGTCTGTAGTTTTGTTAATAGTAATG
CACCAATGTCAGTTGCCTAGTTGTGACAAATATACCGTGGTTATGTAAGATGGTAACATT
AGGGGGAACTGGAGAAGGGTAGATTGGAGCTCTCTGTACTATCTTTGCAACTTTTCTGGG
AATCTAAAATTACTCCAAAATAAAAAAAAAATGTATTTAAAGTAAATATATTCCCTAAGA
GTCCAGGAGGCAGGGGAGTTGTAGAAGCAGCTGAGTGGTTGGGTTCTGACAGATTTGGTT
CCAACTCGGTCTCTGCTGCTCACCAGCTGTGTGACCTTGAGCAAGTGGCTTAGCCTTTCT
GAGCCTGATTTCCTTATCTGTGGAGTGGGGAAGATGACAGCCACCTCGCAGGGCTGTGGA
GGGTTAAACGAGGTGATGCATGGACAGCAGCCGCACTGACCTTGCTGGTGTGGGGCTCCT
GCTTCTGTTCTTCCCGTGCAGCCTTGGGAATGTTGGAGGCCGTATCCAGGGACCCCTGGG
CCTCCTGGGATGGCCTCTCTGGATCAGCCTTGGAAGGTTCCAGGCTGCCCTTAGGCTCCC
ACATTCTTCCCCAGTCACGCTCTCCTCGCCCTGCCCACACCAGTCCTGTGACCCTTGCCT
GAGTTGTGACTTCCCACCCCTCCCCGGCCTAGAGGAAAGCTGCCTGGCCCCTCAGTGGGA
CTCCCGCCCACTGACCCTCTGTCCACCATACACAGACAGGGGCACTATCCACAAGGTGGT
GGAACCGGGGGAGCAGGAGCACAGCTTCGCCTTCAACATCATGGAGATCCAGCCCTTCCG
CCGCGCGGCTGCCATCCAGACCATGTCGCTGGATGCTGAGCGGGTGAGCCTTCCCCCACT
GCGTCCCATGGGCTATGCAGTGACTGCAGCTGAGGACAGGGCTCCTTTGCATGTGATTTG
TGTGTTCTTTTAAGAGCTTCTAGGCCTTAGGGCCTGGACATTTAGGACTGAGTGTGGGGT
GGGGCCCGGGCCTGACCCAATCCTGCTGTCCTTCCAGAGGAAGCTGTATGTGAGCTCCCA
GTGGGAGGTGAGCCAGGTGCCCCTGGACCTGTGTGAGGTCTATGGCGGGGGCTGCCACGG
TTGCCTCATGTCCCGAGACCCCTACTGCGGCTGGGACCAGGGCCGCTGCATCTCCATCTA
CAGCTCCGAACGGTACGTTGGCCGGGATCCCTCCGTCCCTGGGACAAGGTGGGCATGGGA
CAGGGGGAGGTGTTGTCGGGCTGGAAGAGGTGGCGGTACTGGGCCTTTCTTGTGGGACCT
CCTCTCTACTGGAACTGCACTAGGGGTAAGGATATGAGGGTCAGGTCTGCAGCCTTGTAT
CTGCTGATCCTCTTTCGTCCTTCCCACTCCAGGTCAGTGCTGCAATCCATTAATCCAGCC
GAGCCACACAAGGAGTGTCCCAACCCCAAACCAGGTACCTGATCTGGCCCTGCTGGCGGC
TGTGGCCCAATGAGTGGGGTACTGCCCTGCCCTGATTGTCCTGGTCTGAGGGAAACATGG

CCTTGTCCTGTGGGCCCCAGGTACATGGGGCAGGATACAGTCCTGCAGAGGGAGCCCTCT
TGGTGGGATGAGCGAGACGGGAGAAAAAAGGAGGACGCTGAGGGCTGGGTTCCCCACGTT
CATTCAGAAGCCTTGTCCTGGGATCCCAGTCGGTGGGGAGGACACATCCTCCCCTGGGAG
CTCTTTGTCCCTCCTCACGGCTGCTTCCCCACTGCCTCCCCAGACAAGGCCCCACTGCAG
AAGGTTTCCCTGGCCCCAAACTCTCGCTACTACCTGAGCTGCCCCATGGAATCCCGCCAC
GCCACCTACTCATGGCGCCACAAGGAGAACGTGGAGCAGAGCTGCGAACCTGGTCACCAG
AGCCCCAACTGCATCCTGTTCATCGAGAACCTCACGGCGCAGCAGTACGGCCACTACTTC
TGCGAGGCCCAGGAGGGCTCCTACTTCCGCGAGGCTCAGCACTGGCAGCTGCTGCCCGAG
GACGGCATCATGGCCGAGCACCTGCTGGGTCATGCCTGTGCCCTGGCCGCCTCCCTCTGG
CTGGGGGTGCTGCCCACACTCACTCTTGGCTTGCTGGTCCACTAGGGCCTCCCGAGGCTG
GGCATGCCTCAGGCTTCTGCAGCCCAGGGCACTAGAACGTCTCACACTCAGAGCCGGCTG
GCCCGGGAGCTCCTTGCCTGCCACTTCTTCCAGGGGACAGAATAACCCAGTGGAGGATGC
CAGGCCTGGAGACGTCCAGCCGCAGGCGGCTGCTGGGCCCCAGGTGGCGCACGGATGGTG
AGGGGCTGAGAATGAGGGCACCGACTGTGAAGCTGGGGCATCGATGACCCAAGACTTTAT
CTTCTGGAAAATATTTTTCAGACTCCTCAAACTTGACTAAATGCAGCGATGCTCCCAGCC
CAAGAGCCCATGGGTCGGGGAGTGGGTTTGGATAGGAGAGCTGGGACTCCATCTCGACCC
TGGGGCTGAGGCCTGAGTCCTTCTGGACTCTTGGTACCCACATTGCCTCCTTCCCCTCCC
TCTCTCATGGCTGGGTGGCTGGTGTTCCTGAAGACCCAGGGCTACCCTCTGTCCAGCCCT
GTCCTCTGCAGCTCCCTCTCTGGTCCTGGGTCCCACAGGACAGCCGCCTTGCATGTTTAT
TGAAGGATGTTTGCTTTCCGGACGGAAGGACGGAAAAAGCTCTGAAAAAAAAAAAAAAAA
AAAAAAAA

Table15: Nucleotide sequence of pMelBacA-H-SEMAL (6622bp) (SEQ ID NO: 42)

1 GATATCATGG AGATAATTAA AATGATAACC ATCTCGCAAA TAAATAAGTA

51 TTTTACTGTT TTCGTAACAG TTTTGTAATA AAAAAACCTA TAAATATGAA

101 ATTCTTAGTC AACGTTGCCC TTGTTTTTAT GGTCGTATAC ATTTCTTACA

151 TCTATGCGGA TCGATGG

gga tccgcccagg gccacctaag gagcggaccc

201 cgcatcttcg ccgtctggaa aggccatgta gggcaggacc gggtggactt

251 tggccagact gagccgcaca cggtgctttt ccacgagcca ggcagctcct

301 ctgtgtgggt gggaggacgt ggcaaggtct acctctttga cttccccgag

351 ggcaagaacg catctgtgcg cacggtgaat atcggctcca caaaggggtc

401 ctgtctggat aagcgggact gcgagaacta catcactctc ctggagaggc

451 ggagtgaggg gctgctggcc tgtggcacca acgcccggca ccccagctgc

501 tggaacctgg tgaatggcac tgtggtgcca cttggcgaga tgagaggcta

551 tgccccttc agcccggacg agaactccct ggttctgttt gaaggggacg

601 aggtgtattc caccatccgg aagcaggaat acaatgggaa gatccctcgg

651 ttccgccgca tccggggcga gagtgagctg tacaccagtg atactgtcat

701 gcagaaccca cagttcatca aagccaccat cgtgcaccaa gaccaggctt

751 acgatgacaa gatctactac ttcttccgag aggacaatcc tgacaagaat

801 cctgaggctc ctctcaatgt gtcccgtgtg gcccagttgt gcaggggga

851 ccagggtggg gaaagttcac tgtcagtctc caagtggaac acttttctga

901 aagccatgct ggtatgcagt gatgctgcca ccaacaagaa cttcaacagg

951 ctgcaagacg tcttcctgct ccctgacccc agcggccagt ggagggacac

1001 cagggtctat ggtgttttct ccaacccctg gaactactca gccgtctgtg

1051 tgtattccct cggtgacatt gacaaggtct tccgtacctc ctcactcaag

1101 ggctaccact caagccttcc caacccgcgg cctggcaagt gcctcccaga

1151 ccagcagccg atacccacag agaccttcca ggtggctgac cgtcacccag

1201 aggtggcgca gagggtggag cccatggggc ctctgaagac gccattgttc

1251 cactctaaat accactacca gaaagtggcc gttcaccgca tgcaagccag

1301 ccacggggag acctttcatg tgctttacct aactacagac aggggcacta

1351 tccacaaggt ggtggaaccg ggggagcagg agcacagctt cgccttcaac

1401 atcatggaga tccagccctt ccgccgcgcg gctgccatcc agaccatgtc

1451 gctggatgct gagcggagga agctgtatgt gagctcccag tgggaggtga

1501 gccaggtgcc cctggacctg tgtgaggtct atggcggggg ctgccacggt

1551 tgcctcatgt cccgagaccc ctactgcggc tgggaccagg gccgctgcat

1601 ctccatctac agctccgaac ggtcagtgct gcaatccatt aatccagccg

1651 agccacacaa ggagtgtccc aaccccaaac cagacaaggc cccactgcag

1701 aaggtttccc tggccccaaa ctctcgctac tacctgagct gccccatgga

1751 atcccgccac gccacctact catggcgcca caaggagaac gtggagcaga

1801 gctgcgaacc tggtcaccag agccccaact gcatcctgtt catcgagaac

1851 ctcacggcgc agcagtacgg ccactacttc tgcgaggccc aggagggctc

1901 ctacttccgc gaggctcagc actggcagct gctgcccgag gacggcatca

1951 tggccgagca cctgctgggt catgcctgtg ccctggctgc ctgaattc

GA

2001 AGCTTGGAGT CGACTCTGCT GAAGAGGAGG AAATTCTCCT TGAAGTTTCC

2051 CTGGTGTTCA AAGTAAAGGA GTTTGCACCA GACGCACCTC TGTTCACTGG

2101 TCCGGCGTAT TAAAACACGA TACATTGTTA TTAGTACATT TATTAAGCGC

2151 TAGATTCTGT GCGTTGTTGA TTTACAGACA ATTGTTGTAC GTATTTTAAT

2201 AATTCATTAA ATTTATAATC TTTAGGGTGG TATGTTAGAG CGAAAATCAA

2251 ATGATTTTCA GCGTCTTTAT ATCTGAATTT AAATATTAAA TCCTCAATAG

2301 ATTTGTAAAA TAGGTTTCGA TTAGTTTCAA ACAAGGGTTG TTTTTCCGAA

2351 CCGATGGCTG GACTATCTAA TGGATTTTCG CTCAACGCCA CAAAACTTGC

2401 CAAATCTTGT AGCAGCAATC TAGCTTTGTC GATATTCGTT TGTGTTTTGT

2451 TTTGTAATAA AGGTTCGACG TCGTTCAAAA TATTATGCGC TTTTGTATTT

2501 CTTTCATCAC TGTCGTTAGT GTACAATTGA CTCGACGTAA ACACGTTAAA

2551 TAAAGCCTGG ACATATTTAA CATCGGGCGT GTTAGCTTTA TTAGGCCGAT

2601 TATCGTCGTC GTCCCAACCC TCGTCGTTAG AAGTTGCTTC CGAAGACGAT

2651 TTTGCCATAG CCACACGACG CCTATTAATT GTGTCGGCTA ACACGTCCGC

2701 GATCAAATTT GTAGTTGAGC TTTTTGGAAT TATTTCTGAT TGCGGGCGTT

2751 TTTGGGCGGG TTTCAATCTA ACTGTGCCCG ATTTTAATTC AGACAACACG

2801 TTAGAAAGCG ATGGTGCAGG CGGTGGTAAC ATTTCAGACG GCAAATCTAC

2851 TAATGGCGGC GGTGGTGGAG CTGATGATAA ATCTACCATC GGTGGAGGCG

2901 CAGGCGGGGC TGGCGGCGGA GGCGGAGGCG GAGGTGGTGG CGGTGATGCA

2951 GACGGCGGTT TAGGCTCAAA TTGTCTCTTT CAGGCAACAC AGTCGGCACC

3001 TCAACTATTG TACTGGTTTC GGGCGTATGG TGCACTCTCA GTACAATCTG

3051 CTCTGATGCC GCATAGTTAA GCCAGCCCCG ACACCCGCCA ACACCCGCTG

3101 ACGCGCCCTG ACGGGCTTGT CTGCTCCCGG CATCCGCTTA CAGACAAGCT

3151 GTGACCGTCT CCGGGAGCTG CATGTGTCAG AGGTTTTCAC CGTCATCACC

3201 GAAACGCGCG AGACGAAAGG GCCTCGTGAT ACGCCTATTT TTATAGGTTA

3251 ATGTCATGAT AATAATGGTT TCTTAGACGT CAGGTGGCAC TTTTCGGGGA

3301 AATGTGCGCG GAACCCCTAT TTGTTTATTT TTCTAAATAC ATTCAAATAT

3351 GTATCCGCTC ATGAGACAAT AACCCTGATA AATGCTTCAA TAATATTGAA

3401 AAAGGAAGAG TATGAGTATT CAACATTTCC GTGTCGCCCT TATTCCCTTT

3451 TTTGCGGCAT TTTGCCTTCC TGTTTTTGCT CACCCAGAAA CGCTGGTGAA

3501 AGTAAAAGAT GCTGAAGATC AGTTGGGTGC ACGAGTGGGT TACATCGAAC

3551 TGGATCTCAA CAGCGGTAAG ATCCTTGAGA GTTTTCGCCC CGAAGAACGT

3601 TTTCCAATGA TGAGCACTTT TAAAGTTCTG CTATGTGGCG CGGTATTATC

3651 CCGTATTGAC GCCGGGCAAG AGCAACTCGG TCGCCGCATA CACTATTCTC

3701 AGAATGACTT GGTTGAGTAC TCACCAGTCA CAGAAAAGCA TCTTACGGAT

3751 GGCATGACAG TAAGAGAATT ATGCAGTGCT GCCATAACCA TGAGTGATAA

3801 CACTGCGGCC AACTTACTTC TGACAACGAT CGGAGGACCG AAGGAGCTAA

3851 CCGCTTTTTT GCACAACATG GGGGATCATG TAACTCGCCT TGATCGTTGG

3901 GAACCGGAGC TGAATGAAGC CATACCAAAC GACGAGCGTG ACACCACGAT

3951 GCCTGTAGCA ATGGCAACAA CGTTGCGCAA ACTATTAACT GGCGAACTAC

4001 TTACTCTAGC TTCCCGGCAA CAATTAATAG ACTGGATGGA GGCGGATAAA

4051 GTTGCAGGAC CACTTCTGCG CTCGGCCCTT CCGGCTGGCT GGTTTATTGC

4101 TGATAAATCT GGAGCCGGTG AGCGTGGGTC TCGCGGTATC ATTGCAGCAC

4151 TGGGGCCAGA TGGTAAGCCC TCCCGTATCG TAGTTATCTA CACGACGGGG

4201 AGTCAGGCAA CTATGGATGA ACGAAATAGA CAGATCGCTG AGATAGGTGC

4251 CTCACTGATT AAGCATTGGT AACTGTCAGA CCAAGTTTAC TCATATATAC

4301 TTTAGATTGA TTTAAAACTT CATTTTTAAT TTAAAAGGAT CTAGGTGAAG

4351 ATCCTTTTTG ATAATCTCAT GACCAAAATC CCTTAACGTG AGTTTTCGTT

4401 CCACTGAGCG TCAGACCCCG TAGAAAAGAT CAAAGGATCT TCTTGAGATC

4451 CTTTTTTTCT GCGCGTAATC TGCTGCTTGC AAACAAAAAA ACCACCGCTA

4501 CCAGCGGTGG TTTGTTTGCC GGATCAAGAG CTACCAACTC TTTTTCCGAA

4551 GGTAACTGGC TTCAGCAGAG CGCAGATACC AAATACTGTT CTTCTAGTGT

4601 AGCCGTAGTT AGGCCACCAC TTCAAGAACT CTGTAGCACC GCCTACATAC

4651 CTCGCTCTGC TAATCCTGTT ACCAGTGGCT GCTGCCAGTG GCGATAAGTC

4701 GTGTCTTACC GGGTTGGACT CAAGACGATA GTTACCGGAT AAGGCGCAGC

4751 GGTCGGGCTG AACGGGGGGT TCGTGCACAC AGCCCAGCTT GGAGCGAACG

4801 ACCTACACCG AACTGAGATA CCTACAGCGT GAGCTATGAG AAAGCGCCAC

4851 GCTTCCCGAA GGGAGAAAGG CGGACAGGTA TCCGGTAAGC GGCAGGGTCG

4901 GAACAGGAGA GCGCACGAGG GAGCTTCCAG GGGGAAACGC CTGGTATCTT

4951 TATAGTCCTG TCGGGTTTCG CCACCTCTGA CTTGAGCGTC GATTTTTGTG

5001 ATGCTCGTCA GGGGGGCGGA GCCTATGGAA AAACGCCAGC AACGCGGCCT

5051 TTTTACGGTT CCTGGCCTTT TGCTGGCCTT TTGCTCACAT GTTCTTTCCT

5101 GCGTTATCCC CTGATTCTGT GGATAACCGT ATTACCGCCT TTGAGTGAGC

5151 TGATACCGCT CGCCGCAGCC GAACGACCGA GCGCAGCGAG TCAGTGAGCG

5201 AGGAAGCATC CTGCACCATC GTCTGCTCAT CCATGACCTG ACCATGCAGA

5251 GGATGATGCT CGTGACGGTT AACGCCTCGA ATCAGCAACG GCTTGCCGTT

5301 CAGCAGCAGC AGACCATTTT CAATCCGCAC CTCGCGGAAA CCGACATCGC

5351 AGGCTTCTGC TTCAATCAGC GTGCCGTCGG CGGTGTGCAG TTCAACCACC

5401 GCACGATAGA GATTCGGGAT TTCGGCGCTC CACAGTTTCG GGTTTTCGAC

5451 GTTCAGACGT AGTGTGACGC GATCGGTATA ACCACCACGC TCATCGATAA

5501 TTTCACCGCC GAAAGGCGCG GTGCCGCTGG CGACCTGCGT TTCACCCTGC

5551 CATAAAGAAA CTGTTACCCG TAGGTAGTCA CGCAACTCGC CGCACATCTG

5601 AACTTCAGCC TCCAGTACAG CGCGGCTGAA ATCATCATTA AAGCGAGTGG

5651 CAACATGGAA ATCGCTGATT TGTGTAGTCG GTTTATGCAG CAACGAGACG

5701 TCACGGAAAA TGCCGCTCAT CCGCCACATA TCCTGATCTT CCAGATAACT

5751 GCCGTCACTC CAACGCAGCA CCATCACCGC GAGGCGGTTT TCTCCGGCGC

5801 GTAAAAATGC GCTCAGGTCA AATTCAGACG GCAAACGACT GTCCTGGCCG

5851 TAACCGACCC AGCGCCCGTT GCACCACAGA TGAAACGCCG AGTTAACGCC

5901 ATCAAAAATA ATTCGCGTCT GGCCTTCCTG TAGCCAGCTT TCATCAACAT

5951 TAAATGTGAG CGAGTAACAA CCCGTCGGAT TCTCCGTGGG AACAAACGGC

6001 GGATTGACCG TAATGGGATA GGTCACGTTG GTGTAGATGG GCGCATCGTA

6051 ACCGTGCATC TGCCAGTTTG AGGGGACGAC GACAGTATCG GCCTCAGGAA

6101 GATCGCACTC CAGCCAGCTT TCCGGCACCG CTTCTGGTGC CGGAAACCAG

6151 GCAAAGCGCC ATTCGCCATT CAGGCTGCGC AACTGTTGGG AAGGGCGATC

6201 GGTGCGGGCC TCTTCGCTAT TACGCCAGCT GGCGAAAGGG GGATGTGCTG

6251 CAAGGCGATT AAGTTGGGTA ACGCCAGGGT TTTCCCAGTC ACGACGTTGT

6301 AAAACGACGG GATCTATCAT TTTTAGCAGT GATTCTAATT GCAGCTGCTC

6351 TTTGATACAA CTAATTTTAC GACGACGATG CGAGCTTTTA TTCAACCGAG

6401 CGTGCATGTT TGCAATCGTG CAAGCGTTAT CAATTTTTCA TTATCGTATT

6451 GTTGCACATC AACAGGCTGG ACACCACGTT GAACTCGCCG CAGTTTTGCG

6501 GCAAGTTGGA CCCGCCGCGC ATCCAATGCA AACTTTCCGA CATTCTGTTG

6551 CCTACGAACG ATTGATTCTT TGTCCATTGA TCGAAGCGAG TGCCTTCGAC

6601 TTTTTCGTGT CCAGTGTGGC TT

The above description of the invention is intended to be illustrative and not limiting. Various changes or modifications in the embodiments described may occur to those skilled in the art. These can be made without departing from the spirit or scope of the invention. Accordingly, it is intended that the invention be limited only to the extent required by the claims and the applicable rules of law.